

Supporting Information

Using CEST NMR to Discover Previously Unobserved States on the Free Energy Surface of Proteins: Application to the L99A Cavity Mutant of T4 Lysozyme

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Experimental Procedures

Generation of synthetic CPMG and CEST data. To generate synthetic CPMG relaxation dispersion data of the type derived from the constant-time CW-CPMG experiment (1-3), the constant time exchange period T_{EX} was set to 20 ms and ν_{CPMG} ($=1/(4\tau_{CPMG})$) varied from 50 Hz to 1000 Hz, where $2\tau_{CPMG}$ is the time between successive refocusing pulses in the CPMG pulse train. The effective transverse relaxation rate is given by $R_{2,eff}(\nu_{CPMG}) = \left(\frac{-1}{T_{EX}}\right) \ln\left(\frac{I(\nu_{CPMG})}{I_0}\right)$ where $I(\nu_{CPMG})$ is the calculated intensity at the end of the T_{EX} period and I_0 is the intensity in the absence of the T_{EX} period (4). Gaussian random noise corresponding to the higher of 0.3 s⁻¹ or 1.5 % of each $R_{2,eff}$ value was added to the calculated intensities. Synthetic ¹⁵N CEST datasets (5) were generated for two different B_1 values: 17 Hz (T_{EX} = 450 ms) and 35 Hz (T_{EX} = 400 ms), with Gaussian noise corresponding to an uncertainty of 0.3 % in the intensities, I , added to the calculated I values. To construct synthetic ¹⁵N CPMG and CEST datasets, ‘reasonable’ $\Delta\varpi_{AB}$ and $\Delta\varpi_{AC}$ values were obtained from the 58 $\Delta\varpi_{FI1}$ ($\Delta\varpi_{AB}$) and $\Delta\varpi_{FI2}$ ($\Delta\varpi_{AC}$) values generated from fits of CEST data from a previous study focussed on the folding trajectory of the A39G FF domain (6), setting $R_{2,A} = R_{2,B} = R_{2,C} = 10$ s⁻¹ and $R_{1,A} = R_{1,B} = R_{1,C} = 1$ s⁻¹.

Analysis of CEST data. For the two-state analysis of the CEST data recorded on the [U-¹⁵N; ²H; Ile δ 1-¹³CH₃; Leu, Val-¹³CH₃/¹²CD₃] (ILV) L99A T4L sample (11.5 °C), ¹⁵N CEST data from the amide sites of 18 residues (L7, R8, E11, I29, V75, I100, N101, F104, Q105, M106, E108, G110, A112, G113, A134, K135, S136, and T142) with $|\Delta\varpi_{EB}| > 2.0$ ppm, and methyl ¹³C CEST data from methyl sites of 5 residues (V103C γ 2, V111C γ 2, L121C δ 1, L118C δ 2 and L133C δ 2) with $|\Delta\varpi_{EB}| > 1.0$ ppm were fit together. The global fitting parameters were the exchange rate ($k_{ex,EB}$) and the minor (B) state population (p_B), while the site-specific fitting parameters were the E (ϖ_E) and B (ϖ_B) state chemical shifts, the longitudinal relaxation rates of spins in state E ($R_{1,E}$) and the transverse relaxation rates of spins in the E ($R_{2,E}$) and B ($R_{2,B}$) states. In all analyses of CEST data carried out here the longitudinal relaxation rate of a given spin was assumed to be the same in all states. Some of the fits were carried out subject to the constraint $R_{2,E} = R_{2,B}$, as described in the main text.

The three-state analysis of the 11.5 °C CEST data recorded on the [U-¹⁵N; ²H; Ile δ 1-¹³CH₃; Leu, Val-¹³CH₃/¹²CD₃] (ILV) L99A T4L sample was carried out using a protocol we recently described (7). Briefly the (relative) signs of the chemical shift differences between the minor state resonances ($\Delta\varpi_{BI}$) were obtained by analysing the (low B_1) ¹⁵N (L7, R8, E11, I29, N101, M106, G110, A112, S136, T142) and methyl ¹³C (V111C γ 2, L118C δ 2, L121C δ 1) CEST profiles recorded at 6, 11.5, 15 and 20 °C (Figure S2) in a two-state manner. The relative signs of $\Delta\varpi_{BI}$ are then obtained from the temperature dependencies of the fitted two-state $\Delta\varpi$ values ($\Delta\varpi_{2st}$) (Figure S2), with the absolute sign of a given $\Delta\varpi_{BI}$ value assumed to be the same as the

sign of the slope of the corresponding $\Delta\varpi_{2st}$ vs temperature profile. As shown in Figure S2 only the relative signs matter as inverting the signs of all $\Delta\varpi_{BI}$ values simply interchanges states I and B. Only sites for which $\Delta\varpi_{2st}$ monotonically increased or decreased with temperature were included in the three-state analysis, for which the global fitting parameters were the exchange rates ($k_{ex,EB}$, $k_{ex,EI}$, $k_{ex,BI}$) and the minor state populations (p_B , p_I) along with site-specific parameters which included chemical shifts of the three states (ϖ_E , ϖ_B , ϖ_I), and the longitudinal ($R_{1,E}$) and transverse ($R_{2,E}$) relaxation rates of spins in the E state since the relaxation rates of a given spin were assumed to be the same in all three states. Values of ($k_{ex,EB} = 270 \pm 9 \text{ s}^{-1}$, $k_{ex,EI} = 313 \pm 45 \text{ s}^{-1}$, $k_{ex,BI} = 1645 \pm 62 \text{ s}^{-1}$, $p_B = 1.46 \pm 0.01 \%$, and $p_I = 0.28 \pm 0.01 \%$; $\chi_{red}^2 = 1$) were obtained, with the sign of $\Delta\varpi_{BI}$ constrained to that obtained from the two-state analysis of CEST data over the 6 to 20 °C temperature range (Figure S2). The ^{15}N CEST profiles of N140 and the methyl ^{13}C CEST profiles of I150 Cδ1 recorded using high B_I values (51.8, 72.6, and 129.6 Hz for ^{15}N ; 49.1 and 98.2 Hz for ^{13}C) contained broad dips consistent with the I state. When these profiles were included in the analysis excellent quality fits were obtained overall, with exchange parameters similar to those reported above ($\chi_{red}^2 = 1$; $k_{ex,EB} = 297 \pm 6 \text{ s}^{-1}$, $k_{ex,EI} = 170 \pm 37 \text{ s}^{-1}$, $k_{ex,BI} = 1667 \pm 44 \text{ s}^{-1}$, $p_B = 1.5 \pm 0.01 \%$, and $p_I = 0.22 \pm 0.01 \%$). Simpler linear three-state models were also fit to the data (see main text), obtained by setting the appropriate exchange rate to 0 s^{-1} .

In the analysis of ^{15}N CEST profiles from $[\text{U-}^{15}\text{N}]$ samples that contained urea or TFE, $\Delta\varpi_{EI}$ and $\Delta\varpi_{EB}$ values were initialised to those obtained above (Table S4). The three-state analysis of the ^{15}N CEST data from the sample containing 8% TFE resulted in good quality fits ($\chi_{red}^2 = 1.16$) with $k_{ex,EB} = 648 \pm 47 \text{ s}^{-1}$, $k_{ex,EI} = 160 \pm 52 \text{ s}^{-1}$, $k_{ex,BI} = 4110 \pm 172 \text{ s}^{-1}$, $p_B = 2.2 \pm 0.1 \%$, and $p_I = 1.8 \pm 0.1 \%$. In the analysis of ^{15}N CEST profiles to obtain urea m -values, the CEST data at all the urea concentrations were simultaneously fitted assuming that $\Delta\varpi_{EB}$ and $\Delta\varpi_{EI}$ do not depend on urea concentration (8), while ϖ_E was allowed to vary with urea concentration. This procedure resulted in good quality fits ($\chi_{red}^2 = 1$), with the exchange parameters reported in Table S5.

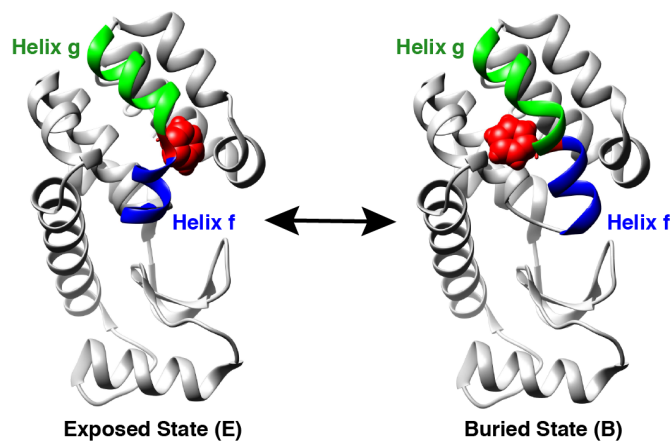


Figure S1 L99A T4L exchanges between a major state (Exposed state) in which Phe114 is exposed to solvent (PDB: 3DMV; (9)) and a minor state (Buried state) in which Phe114 is buried in the core of the protein (PDB: 2LCB; (10)). Phe114 (red) is shown using the CPK representation. In the buried state helix f (blue) and helix g (green) merge to form an extended helix.

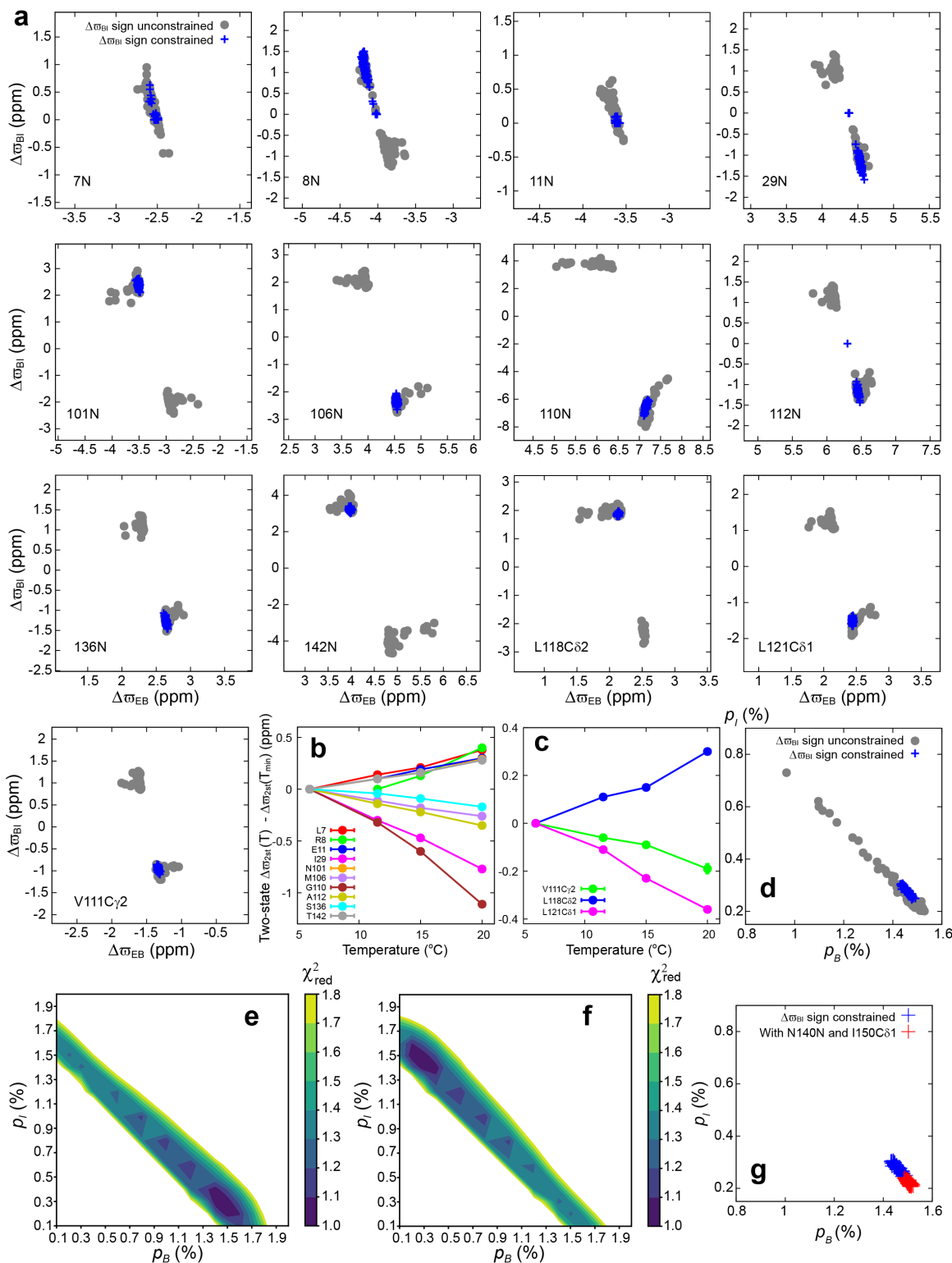


Figure S2 A unique L99A T4L three-state exchange model from amide ^{15}N and methyl ^{13}C CEST data. **(a)** $\Delta\omega_{BI}$ and $\Delta\omega_{EB}$ values for thirteen residues obtained from 100 bootstrap trials from the analysis of CEST data with (blue) and without (grey) constraining the sign of $\Delta\omega_{BI}$. In the absence of constraints $\Delta\omega_{BI}$ can adopt two values of approximately equal magnitude. Signs of $\Delta\omega_{BI}$ are obtained from the temperature dependencies of two-state $\Delta\omega$ values ($\Delta\omega_{2st}$) **(b,c)**. Assuming that $\Delta\omega_{BI}$ values have the same signs as the slopes of the $\Delta\omega_{2st}$ vs temperature plot (on a per-residue basis) leads to a consistent set of relative $\Delta\omega_{BI}$ signs. **(d)** Including sign constraints on $\Delta\omega_{BI}$ results in well defined p_B and p_I values (blue) while in the absence of

constraints p_B and p_I are not defined (grey). **(e,f)** χ^2 surface as function of p_B and p_I obtained by analysing the CEST data using a three-state triangular model. In **(e)** signs are as described above, while in **(f)** the signs of $\Delta\omega_{BI}$ have been inverted i.e., sites for which the slope of the $\Delta\omega_{2st}$ vs temperature plot is +ve have -ve $\Delta\omega_{BI}$ values and vice versa. Equally good fits are obtained when the signs of $\Delta\omega_{BI}$ are inverted, because the optimisation procedure interchanges the labels B and I (as expected), as seen from the fact that the plots in **(e)** and **(f)** are reflected about the diagonal. In **(e)** there is a global minimum with $p_B \gg p_I$. **(g)** Including amide ^{15}N CEST profiles from N140 and $^{13}\text{CH}_3$ methyl CEST data from I150C δ 1 (where the I state dip is visible) in the three-state analysis also results in a minimum (red) consistent with that obtained previously (blue) from **(d)**.

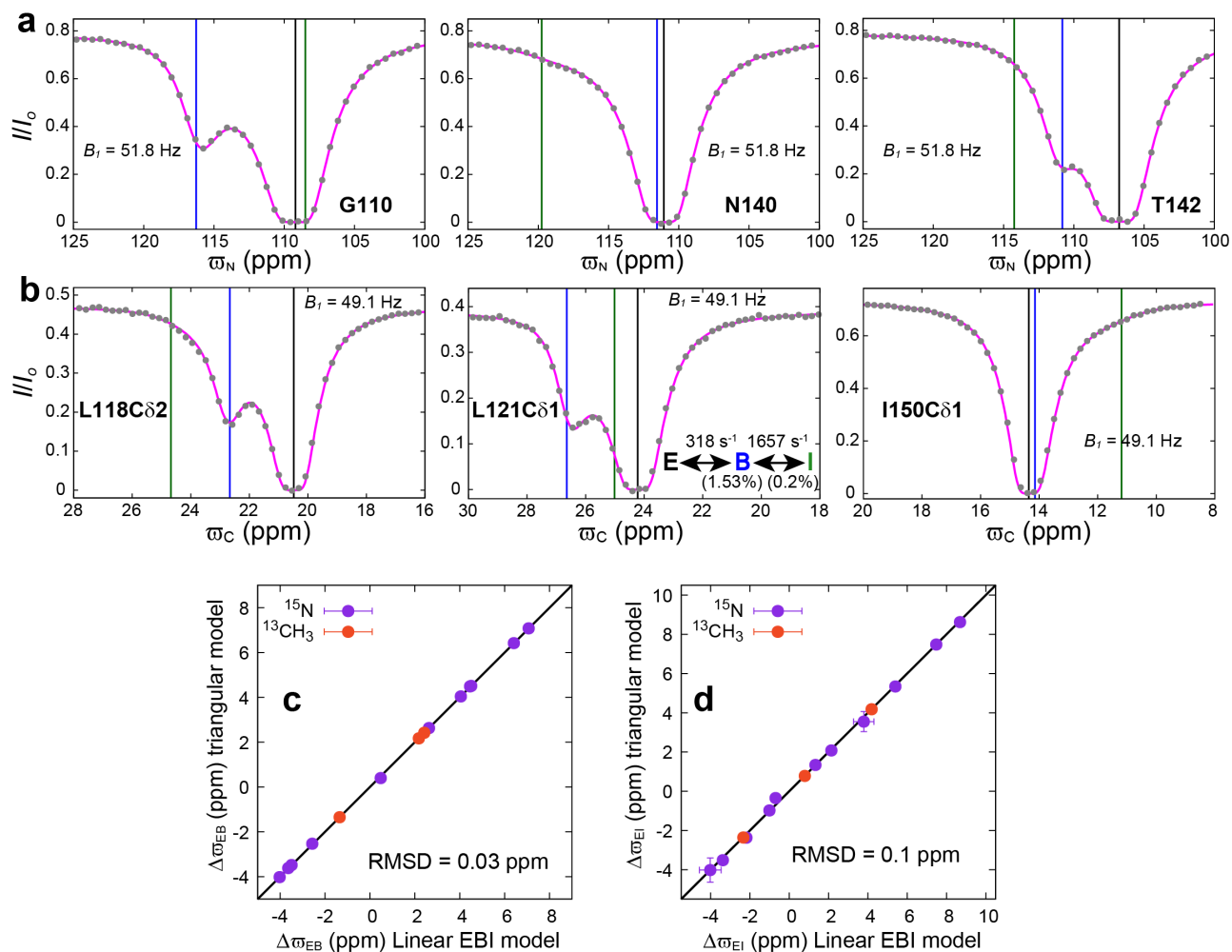


Figure S3 Linear (E↔B↔I) three-state analysis of the L99A T4L CEST data. Representative amide ^{15}N CEST profiles from G110, N140, and T142 **(a)** and methyl ^{13}C CEST profiles from L118C δ 2, L121C δ 1, and I150C δ 1 **(b)**. Experimental data points are shown using grey coloured circles while the magenta line was calculated using the best-fit parameters shown in the central panel in **(b)**. The uncertainties in the I/I_0 values are smaller than the size of the filled circles. $\Delta\omega_{EB}$ **(c)** and $\Delta\omega_{EI}$ **(d)** values obtained from the E↔B↔I model are in excellent agreement with those obtained from the triangular model.

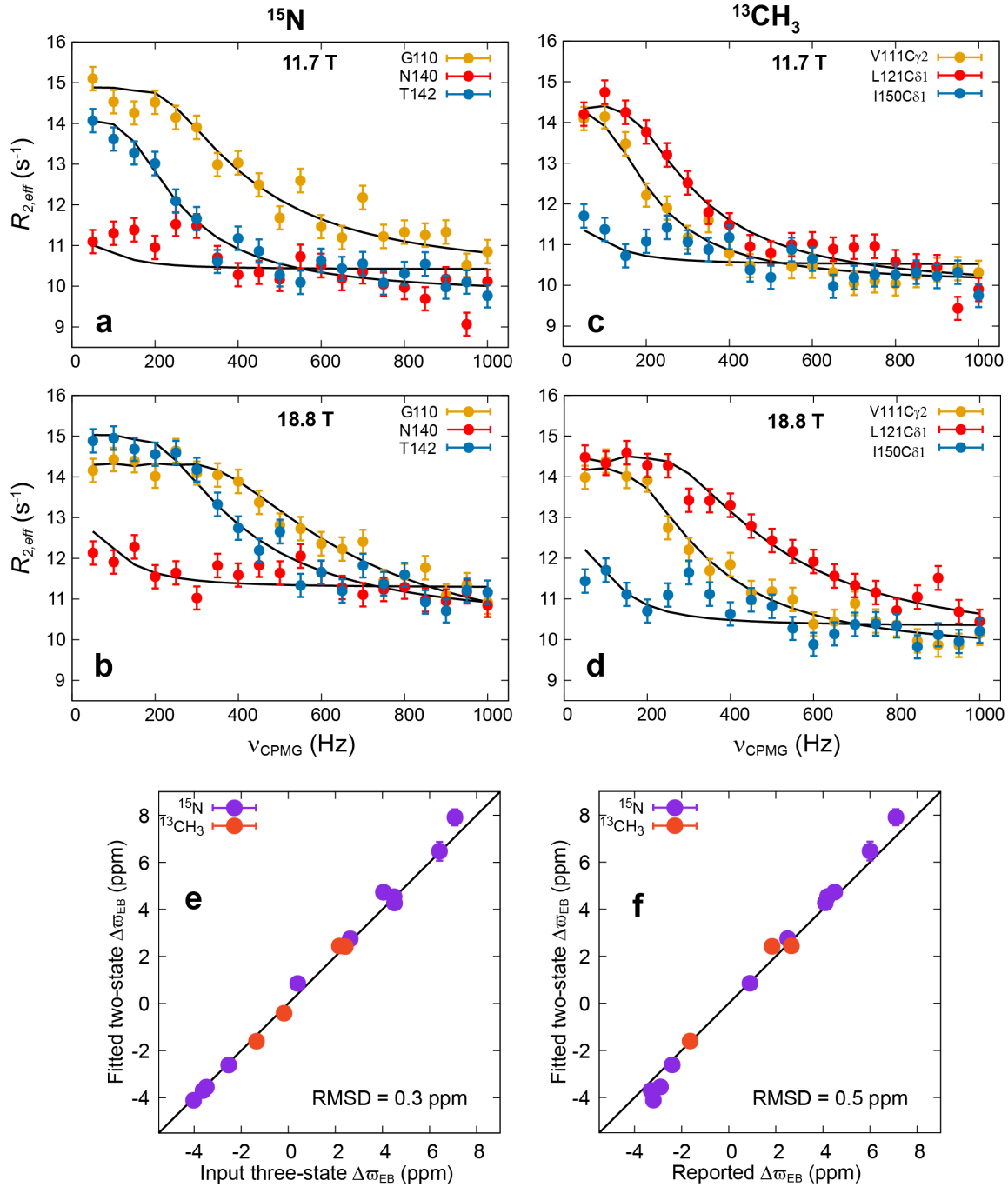


Figure S4 CPMG relaxation dispersion curves are insensitive to the presence of the I state. A two-state exchange model (black line) accounts ($\chi^2_{\text{red}} = 1.04$) for synthetic three-state ^{15}N (a, b) and methyl ^{13}C CPMG (c, d) data. The synthetic data were generated using the best-fit three-state (triangular) exchange parameters ($k_{\text{ex},EB} = 297 \text{ s}^{-1}$, $k_{\text{ex},EI} = 170 \text{ s}^{-1}$, $k_{\text{ex},BI} = 1667 \text{ s}^{-1}$, $p_B = 1.5 \%$, and $p_I = 0.22 \%$) and $\Delta\omega_{EB}$ and $\Delta\omega_{EI}$ values obtained from combined fits of ^{15}N and ^{13}C CEST data (Figure 3). Synthetic CPMG data points are represented using coloured circles while the two-state best-fits ($k_{\text{ex},EB} = 318 \pm 45 \text{ s}^{-1}$ and $p_B = 1.5 \pm 0.3 \%$) are shown with black lines. (e, f) $\Delta\omega_{EB}$ values obtained from two-state fits of the CPMG profiles agree well with the input $\Delta\omega_{EB}$ values used to generate the CPMG profiles (e) as well as with those (Reported $\Delta\omega_{EB}$) values previously obtained (10) from CPMG experiments (f). As the shape of the CPMG relaxation dispersion curve is insensitive to the sign of the fitted $\Delta\omega$ value (11,12), the sign of the fitted $\Delta\omega$ values (Fitted two-state $\Delta\omega_{EB}$) were assumed to be the same as those obtained from the CEST data analysis.

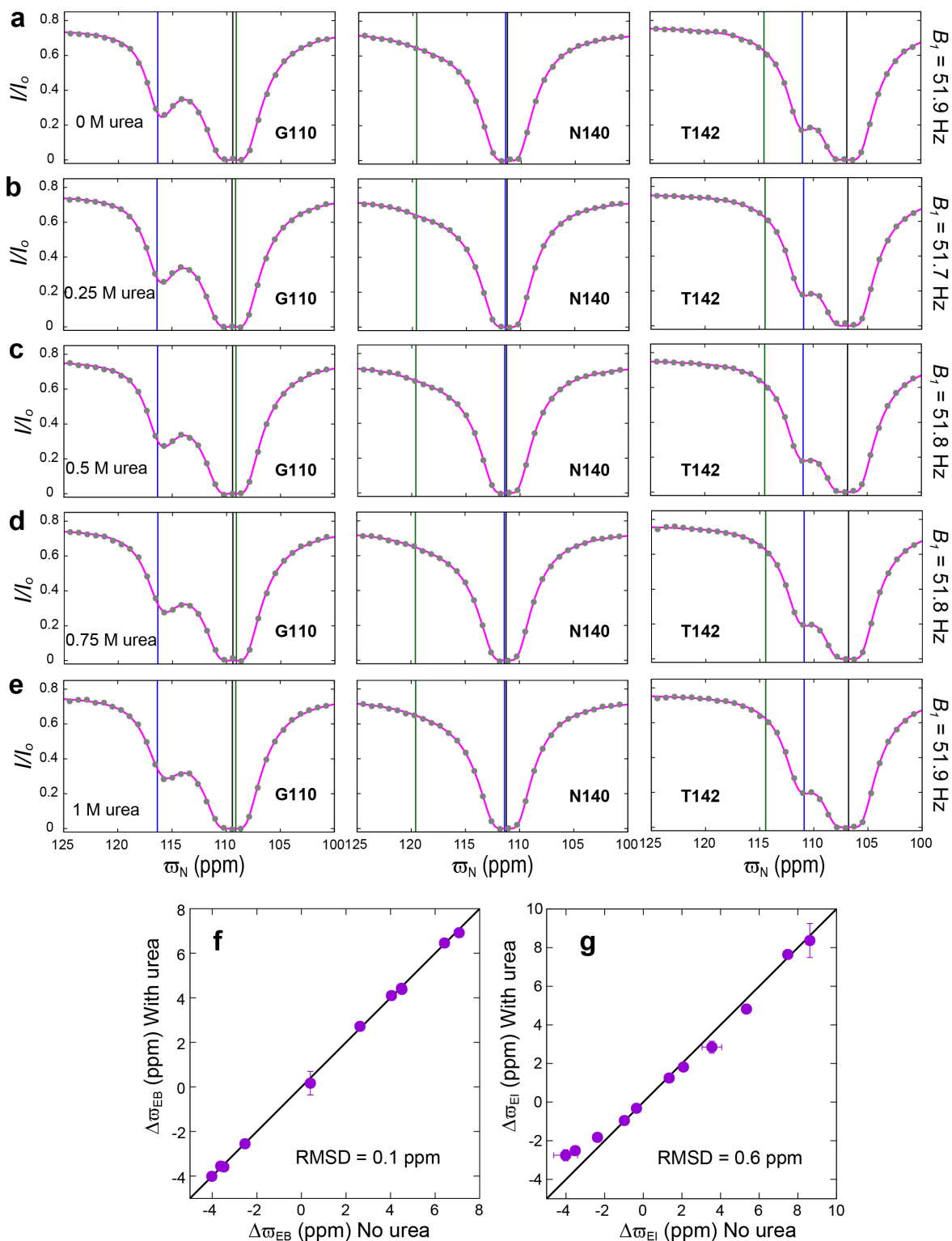


Figure S5 ^{15}N CEST data recorded at various urea concentrations to obtain urea m -values (11.5 °C). Representative ^{15}N CEST data and three-state fits (from residues G110, N140 and T142) recorded using [U- ^{15}N] L99A T4L samples containing 0.0 (a), 0.25 M (b), 0.5 M (c), 0.75 M (d) and 1.0 M (e) urea. The experimental data points are shown as grey circles while the magenta line was calculated using the (three-state) best-fit parameters (Table S5). The uncertainties in the I/I_0 values are smaller than the size of the filled circles. The vertical lines are drawn at the best-fit ω_E (black), ω_B (blue), and ω_I (green) values obtained from

the three-state fits. Data collected at all the urea concentrations were analysed simultaneously with $\Delta\varpi_{EB}$ and $\Delta\varpi_{EI}$ assumed to be urea concentration independent while ϖ_E was allowed to vary with urea concentration. Comparison of amide ^{15}N $\Delta\varpi_{EB}$ (**f**) and $\Delta\varpi_{EI}$ (**g**) values obtained in the analysis of the urea concentration series with those obtained by analysing ^{15}N and ^{13}C CEST data in the absence of urea.

Temperature (°C)	Experiment	B_1 (Hz)	T_{EX} (ms)	Range (Hz)	Step-size (Hz)	Carrier (ppm)
11.5	^{15}N CEST	18.1	450	± 1260	20	119.707
		33.2	450	± 1250	25	119.707
		51.8	400	± 1400	40	119.707
		72.6	375	± 1400	50	119.707
		129.6	350	± 1760	55	119.707
	^{13}C CEST	16.2	475	-1500 to 2000	20	17.868
		32.4	450	-1620 to 2130	30	17.868
		49.1	425	-1650 to 2150	40	17.868
		98.2	375	-1800 to 2260	70	17.868
6.0	^{15}N CEST	31.2	450	± 1260	30	119.765
		67.5	375	± 1400	50	119.765
	^{13}C CEST	27.1	450	-1620 to 2130	30	17.927
15.0	^{15}N CEST	25.9	450	± 1350	25	119.682
		51.8	400	± 1400	40	119.682
	^{13}C CEST	48.9	400	-1650 to 2150	40	17.846
20.0	^{15}N CEST	36.5	330	± 1350	30	119.627
		52.1	325	± 1400	40	119.627
	^{13}C CEST	49.7	340	-1650 to 2150	40	17.789

Table S1 Details of amide ^{15}N and methyl ^{13}C CEST experiments(5,13) using the sample containing ~ 1.2 mM $[\text{U-}^{15}\text{N}; ^2\text{H}; \text{Ile}\delta 1\text{-}^{13}\text{CH}_3; \text{Leu, Val-}^{13}\text{CH}_3/^{12}\text{CD}_3]$ (ILV) L99A T4L dissolved in 50 mM sodium phosphate, 25 mM NaCl, 2 mM NaN_3 , 2 mM EDTA, pH 5.5, (10% D_2O) buffer. The B_1 irradiation was applied for a time T_{EX} over a range of offsets around the carrier, as indicated (Range), and Step-size is the spacing between adjacent offsets. A reference plane without the T_{EX} delay was also recorded. The 11.5 °C datasets were analysed using both two and three-state exchange models while the 6.0, 15.0 and 20.0 °C datasets along with the 11.5 °C dataset were analysed using a two-state model to obtain the signs of $\Delta\varpi_{BI}$ values.

Temperature (°C)	Experiment	B_1 (Hz)	T_{EX} (ms)	Range (Hz)	Step-size (Hz)	Carrier
15	^{15}N CEST	17.5	450	± 1190	20	119.689
15	^{15}N CEST	51.4	350	± 1400	40	119.689
15	^{15}N CEST	102.9	300	± 1760	55	119.689

Table S2 Details of the amide ^{15}N CEST experiments (5) carried out to establish the robustness of the CEST derived ϖ_I values. The sample contained ~ 1.5 mM $[\text{U-}^{15}\text{N}]$ L99A T4L dissolved in 50 mM sodium phosphate, 25 mM NaCl, 2 mM NaN_3 , 2 mM EDTA, 8% (v/v) TFE, pH 5.5, (5% D_2O) buffer. During the exchange time, T_{EX} , B_1 irradiation was applied over a range of offsets around the carrier, with a spacing given by Step-size. A reference plane without the T_{EX} delay was also recorded.

Temperature (°C)	Experiment	Urea Concentration (M)	B_1 (Hz)	T_{EX} (ms)	Range (Hz)	Step-size (Hz)	Carrier
11.5	^{15}N CEST	0.0	26.0	480	± 1155	35	119.706
			51.9	450	± 1400	56	119.706
			114.2	350	± 1680	70	119.706
11.5	^{15}N CEST	0.25	25.9	480	± 1155	35	119.706
			51.7	450	± 1400	56	119.706
			113.7	350	± 1680	70	119.706
11.5	^{15}N CEST	0.5	25.9	480	± 1155	35	119.706
			51.8	450	± 1400	56	119.706
			114.0	350	± 1680	70	119.706
11.5	^{15}N CEST	0.75	26.0	480	± 1155	35	119.706
			51.9	450	± 1400	56	119.706
			114.2	350	± 1680	70	119.706
11.5	^{15}N CEST	1.0	25.9	480	± 1155	35	119.706
			51.8	450	± 1400	56	119.706
			114.0	350	± 1680	70	119.706

Table S3 Details of amide ^{15}N CEST experiments (5) carried out to derive urea m -values for E, B, and I, and the associated transition states using the three-state triangular exchange model. Samples were comprised of ~ 1.5 mM $[\text{U-}^{15}\text{N}]$ L99A T4L dissolved in 50 mM sodium phosphate, 25 mM NaCl, 2 mM NaN_3 , 2 mM EDTA, pH 5.5, (5% D_2O) buffer with an amount of urea as indicated. B_1 irradiation was applied during the exchange time, T_{EX} , over a range of offsets around the carrier, spaced at intervals of Step-size. A reference plane without the T_{EX} delay was also recorded.

Residue	Site	ϖ_E (ppm)	$\Delta\varpi_{EB}$ (ppm)		$\Delta\varpi_{EI}$ (ppm)	
			Value	Uncertainty	Value	Uncertainty
V111	Methyl $^{13}\text{C}\gamma_2$	24.28	-1.35	0.20	-2.36	0.20
L118	Methyl $^{13}\text{C}\delta_2$	20.48	2.17	0.20	4.18	0.20
L121	Methyl $^{13}\text{C}\delta_1$	24.22	2.42	0.20	0.78	0.20
I150	Methyl $^{13}\text{C}\delta_1$	14.35	-0.18	0.20	-3.14	0.23
L7	Amide ^{15}N	118.93	-2.53	0.20	-2.37	0.20
R8	Amide ^{15}N	124.22	-4.02	0.20	-4.02	0.62
E11	Amide ^{15}N	118.78	-3.62	0.20	-3.52	0.20
I29	Amide ^{15}N	129.78	4.49	0.20	3.55	0.51
N101	Amide ^{15}N	119.36	-3.48	0.20	-0.98	0.20
M106	Amide ^{15}N	113.12	4.51	0.20	2.08	0.20
G110	Amide ^{15}N	109.21	7.08	0.20	-0.35	0.22
A112	Amide ^{15}N	117.90	6.42	0.20	5.34	0.20
S136	Amide ^{15}N	113.08	2.63	0.20	1.34	0.20
N140	Amide ^{15}N	111.06	0.40	0.20	8.62	0.20
T142	Amide ^{15}N	106.77	4.04	0.20	7.48	0.20

Table S4 ϖ_E , $\Delta\varpi_{EB}$ and $\Delta\varpi_{EI}$ values obtained by analysing the 11.5 °C CEST data from the ~1.2 mM [U- ^{15}N ; ^2H ; Ile δ_1 - $^{13}\text{CH}_3$; Leu, Val- $^{13}\text{CH}_3$ / $^{12}\text{CD}_3$] (ILV) L99A T4L sample (50 mM sodium phosphate, 25 mM NaCl, 2 mM NaN_3 , 2 mM EDTA, pH 5.5, 10% D_2O , buffer). A triangular three-state model was used. Uncertainties were estimated using a bootstrap procedure (14,15) with 100 trials. The minimum value of the reported uncertainty in $\Delta\varpi_{EB}$ and $\Delta\varpi_{EI}$ was set to 0.2 ppm.

Urea Concentration [M]	$k_{ex,EB}$ (s ⁻¹)	$k_{ex,EI}$ (s ⁻¹)	$k_{ex,BI}$ (s ⁻¹)	p_B (%)	p_I (%)
0.0	236.9 ± 9	355.1 ± 65	1464.9 ± 55	1.57 ± 0.02	0.24 ± 0.01
0.25	265.9 ± 8	297.0 ± 55	1607.5 ± 49	1.56 ± 0.01	0.26 ± 0.01
0.5	283.8 ± 9	285.3 ± 54	1799.1 ± 58	1.49 ± 0.01	0.26 ± 0.01
0.75	303.4 ± 12	257.8 ± 65	2006.4 ± 66	1.43 ± 0.02	0.28 ± 0.02
1.0	322.4 ± 18	303.2 ± 88	2204.3 ± 64	1.36 ± 0.02	0.28 ± 0.02

Table S5 Exchange parameters obtained from a triangular three-state analysis of amide ¹⁵N CEST data recorded at 11.5 °C (Table S3) using ~1.5 mM [U-¹⁵N] L99A T4L samples dissolved in 50 mM sodium phosphate, 25 mM NaCl, 2 mM NaN₃, 2 mM EDTA, pH 5.5, (5% D₂O) buffer, with the amount of urea indicated. ¹⁵N CEST data at all the urea concentrations were simultaneously analysed assuming that $\Delta\varpi_{EB}$ and $\Delta\varpi_{EI}$ are invariant to the addition of urea, while ϖ_E was allowed to vary with urea concentration. Uncertainties were estimated using a bootstrap procedure with 50 trials.

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