



Supporting Information

Enhancing the Sensitivity of CPMG Relaxation Dispersion to Conformational Exchange Processes by Multiple-Quantum Spectroscopy

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Supporting Information

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Materials and Methods

Details of TQ-CPMG pulse sequence. 90° (180°) rectangular pulses, denoted by narrow (wide) bars, are applied with a 25 kHz field between points A and B, and at maximum power elsewhere. All pulses are applied along the x-axis unless otherwise indicated. The hatched bars denote composite 180° pulses^[1], as described below, while the water-selective shaped pulse marked with “w” (~ 7 ms) is implemented using the EBURP-1 profile^[2]. Note that a single 180_{ϕ_2} pulse is applied in the center of the CPMG scheme that decreases the effects of pulse imperfections, as described previously^[3]. Between points A and B the ^1H carrier is placed in the center of the methyl region (~ 1 ppm), while at point B the carrier is positioned at the water resonance (~ 4.7 ppm); ^{13}C and ^2H carriers are at ~ 20 ppm and ~ 1 ppm, respectively. ^{13}C and ^2H WALTZ-16 decoupling elements^[4] are applied with fields of 2 kHz and 500 Hz, respectively. The delays used are: $\tau_a = 1.8$ ms, $\tau_b = 1.8\text{--}2$ ms, $\tau_{cp} = T_{relax}/(4N)$, N is an integer. The phase cycle is $\phi_1 = (0^\circ, 60^\circ, 120^\circ, 180^\circ, 240^\circ, 300^\circ)$; $\phi_2 = y, -y$; $\phi_3 = 6(x), 6(-x)$; $\phi_4 = x$; $\phi_5 = x, -x$; $\phi_{rec} = 3(x, -x), 3(-x, x)$. Typically we record the complete phase cycle for H_2O samples, as the ^{13}C 90° pulse of phase ϕ_3 significantly decreases the residual water signal, while for D_2O samples a 6-step cycle is used ($\phi_3 = x$) so long as sensitivity is not limiting. The phases ψ_1/ψ_2 are used to implement the XY-16 scheme^[5]. Phase ψ_1 is incremented with the cycle $(x, y, x, y, y, x, y, x, -x, -y, -x, -y, -y, -x, -y, -x)$ for each successive N value, with ψ_2 decremented in the same manner, but inverted. Thus, the CPMG element becomes $\tau_{cp}\Pi_1\tau_{cp}\tau_{cp}\Pi_2\tau_{cp}\dots\tau_{cp}\Pi_{N-1}\tau_{cp}\tau_{cp}\Pi_N\tau_{cp}180_{\phi_2}\tau_{cp}\bar{\Pi}_N\tau_{cp}\bar{\Pi}_{N-1}\tau_{cp}\dots\tau_{cp}\bar{\Pi}_2\tau_{cp}\tau_{cp}\bar{\Pi}_1\tau_{cp}$ where $\Pi_1 = \Pi_x, \Pi_2 = \Pi_y, \Pi_3 = \Pi_x$ and so on following the XY-16 scheme, with $\Pi_x = 90_{-y}180_x90_{-y}, \Pi_y = 90_x180_y90_x, \Pi_{-x} = 90_y180_{-x}90_y, \Pi_{-y} = 90_{-x}180_{-y}90_{-x}$. $\bar{\Pi}_j$ values are calculated from Π_j by interchanging $y(-y)$ with $-y(y)$ in expressions for Π_j , that follows from the 180_{ϕ_2} pulse in the center of the pulse scheme as discussed by Hansen et al.^[3] Quadrature detection in F_1 is achieved by STATES-TPPI of ϕ_4 ^[6]. Gradients are applied with the following durations (ms) and strengths (G/cm): g_0 : (1.0, 4), g_1 : (0.4, 5), g_2 : (0.6, 6), g_3 : (0.3, -5), g_4 : (0.5, 10), g_5 : (0.3, -8).

Sample preparation. All proteins were expressed in *E. coli* BL21(DE3) cells grown in M9 minimal media (\sim 100% D₂O), containing ¹⁵N-ammonium chloride (1 g/L) and [²H,¹²C]-glucose (3 g/L) as the sole nitrogen and carbon sources, respectively. Specifically labeled precursors^[7] were added 1 h prior to the induction of protein overexpression with 1 mM IPTG, as described previously^[7,8]. Purification of samples followed literature protocols. In brief, the following samples were used: (i) 1.5 mM [U-²H; Ile δ 1-¹³CH₃; Leu,Val-¹³CH₃/¹²CD₃; Met-¹³CH₃]-labeled B1 domain of immunoglobulin binding protein G^[9] in 50 mM potassium phosphate pH 7.5, 100 mM NaCl, 0.1 mM NaN₃, 100% D₂O; (ii) 1.5 mM [U-²H; Ile δ 1-¹³CH₃; Leu,Val-¹³CH₃/¹²CD₃; Met-¹³CH₃]- and 1.0 mM [U-²H; Ile δ 1-¹³CHD₂; Leu,Val-¹³CHD₂/¹³CHD₂; Met-¹³CHD₂]-labeled FF domain from human HYPA/FBP11^[10] in 25 mM potassium phosphate pH 6.8, 50 mM NaCl, 1 mM EDTA, 100% D₂O; (iii) 0.9 mM (monomer concentration) [U-²H; Ile δ 1-¹³CH₃; Leu,Val-¹³CH₃/¹²CD₃; Met-¹³CH₃]-labeled $\alpha_7\alpha_7$, from *T. acidophilum*^[11], 25 mM potassium phosphate pH 7.4, 50 mM NaCl, 4.6 mM NaN₃, 1 mM EDTA, 100% D₂O. The level of deuteration in all samples was $> 95\%$ at all carbon sites.

NMR spectroscopy. All NMR experiments were recorded on Varian 600 MHz and 800 MHz spectrometers equipped with triple-axis gradient room temperature probes. Data sets were measured at 25°C with the exception of $\alpha_7\alpha_7$ where dispersion profiles were recorded at 50°C. ¹H SQ ¹³CH₃ and ¹³CHD₂ relaxation dispersion data sets were recorded as described in the literature^[12,13], while TQ profiles were measured using the pulse scheme described in the text. ¹H relaxation dispersion experiments have been recorded using a constant-time CPMG element^[14,15] $T_{relax} = 40$ ms (GB1) or 20 ms (FF domain and $\alpha_7\alpha_7$). Experiments were recorded as pseudo-3D datasets by varying the number of CPMG pulses N during T_{relax} for each 2D plane, with the value of T_{relax} chosen such that peak intensities decay to $\sim 50\%$ of their values for $T_{relax} = 0$. Each 2D data set was recorded with 6 scans/FID, a relaxation delay of 2.0 s and (576, 64) complex points in (t_2 , t_1) to give a net acquisition time of ~ 30 min/spectrum. A series of 2D maps was obtained with ν_{CPMG} values varying between 25 Hz ($T_{relax} = 40$ ms)

or 50 Hz ($T_{relax} = 20$ ms) and 2000 Hz. Approximately 30–40 planes were recorded for each dispersion series, including duplicates for error analysis^[16]. The net measurement time for each experiment was ~ 15 –20 h.

Data analysis. All NMR spectra were processed and analyzed using the NMRPipe suite of programs^[17]. Effective transverse relaxation rates, $R_{2,eff}$, were calculated based on peak intensities according to the relation $R_{2,eff}(\nu_{CPMG}) = -\ln(I(\nu_{CPMG})/I_0)/T_{relax}$, where I_0 is the peak intensity in a reference spectrum recorded without the relaxation delay, T_{relax} ^[15]. Fitting of cross-peaks in CPMG data sets was carried out using the software package ChemEx (<https://github.com/gbouvignies/chemex>); a separate module is required for fitting TQ ^1H relaxation dispersion data, which is available upon request. Only residues with TQ(SQ) dispersion profiles such that $R_{ex} = R_{2,eff}(\nu_{CPMG} = 50 \text{ Hz}) - R_{2,eff}(\nu_{CPMG} = 2000 \text{ Hz}) > 7 \text{ s}^{-1}$ (3 s^{-1}) at 800 MHz, were included in the analyses. Exchange parameters were extracted from fits of dispersion data to a two-site exchange model using the Bloch–McConnell equations^[18], as described below. The fitting parameters include the global values p_E and k_{ex} (or alternatively $k_{EG} = (1 - p_E)k_{ex}$ and $k_{GE} = p_Ek_{ex}$, see below) and residue specific chemical shift differences, $\Delta\varpi$ (ppm), and intrinsic relaxation rates for the anti-phase TQ coherence $3C_zI_+^1I_+^2I_-^3 + 3C_zI_-^1I_-^2I_+^3$.

As described in the main text, the resulting imbalance between the relaxation rates of in-phase and anti-phase coherences^[19], that interconvert during intervals between ^1H CPMG refocusing pulses, can lead to small contributions to $R_{2,eff}$ values in dispersion profiles that can be corrected so long as the difference in transverse relaxation rates of these coherences is taken into account. The differential relaxation between in-phase and anti-phase TQ coherences, $\Delta R_2 = R_{2,I} - R_{2,A}$, can be measured with the scheme shown in Figure S1 (see below) with ΔR_2 subsequently inserted as a fixed parameter in the fits or, alternatively, ΔR_2 can be treated as a fitting parameter for each residue. The two approaches yield essentially identical results for the FF domain and we prefer, therefore, treating ΔR_2 as a fitting parameter as this involves less measurements.

Further discussion of the sensitivity of the TQ CPMG pulse scheme. In the text we have presented a brief explanation as to why 3/4 of the signal is retained in the TQ experiment. It is important to note that the terms of interest at the start of the CPMG element ($8C_zI_x^1I_y^2I_x^3 + 8C_zI_y^1I_x^2I_y^3 + 8C_zI_y^1I_y^2I_x^3 = 8C_zQ$) are subsequently selected at the end of T_{relax} and transferred back to observable ^1H magnetization. This gives rise to the factor of 3/4 discussed in the text. Instead, imagine an experiment where the magnetization of interest at the start of the CPMG element is given by $8C_zI_x^1I_x^2I_x^3$. As with $8C_zQ$ it is straightforward to show that $8C_zI_x^1I_x^2I_x^3$ comprises both TQ and SQ coherences. Selection of the TQ elements results in the following terms, $8C_zI_x^1I_x^2I_x^3 \xrightarrow{\text{TQ selection}} C_zI_+^1I_+^2I_+^3 + C_zI_-^1I_-^2I_-^3$. If the pulse scheme subsequently selects for elements proportional to $8C_zI_x^1I_x^2I_x^3$ at end of the CPMG interval then only 1/4 of the starting coherences will ultimately be detected by observing ^1H . Thus, the ‘nature’ of the coherences from which the TQ elements are selected and subsequently returned to is critical in defining the sensitivity of the experiment.

Differential relaxation between anti-phase and in-phase ^1H TQ elements, ΔR_2 . It is of interest to establish the magnitude of $\Delta R_2 = R_{2,I} - R_{2,A}$ and in particular how ΔR_2 relates to the methyl ^{13}C R_1 rate, as ΔR_2 derives from differences in relaxation rates of coherences that are in and out of phase with respect to ^{13}C . A Redfield density matrix calculation^[20] shows that

$$\Delta R_2 = 0.3d_{CH}^2[3J(\omega_C) + 6K(\omega_C)] \quad [\text{S1}]$$

where $d_{CH}^2 = \left(\frac{\mu_0}{4\pi}\right)^2 \frac{\gamma_H^2 \gamma_C^2 \hbar^2}{r_{HC}^6}$, γ_j is the gyromagnetic ratio of spin j , h is Planck’s constant, and r_{HC} is the one-bond ^1H – ^{13}C distance. In Eq. [S1] $J(\omega)$ and $K(\omega)$ are auto- and cross-correlation spectral densities for the ^{13}C – ^1H dipolar interactions, defined as^[21,22]:

$$J(\omega) = \frac{1}{9}S_{axis}^2 \frac{\tau_c}{1 + (\omega\tau_c)^2} + \left(1 - \frac{1}{9}S_{axis}^2\right) \frac{\tau'}{1 + (\omega\tau')^2}, \quad [\text{S2}]$$

$$K(\omega) = \frac{1}{9}S_{axis}^2 \frac{\tau_c}{1 + (\omega\tau_c)^2} + \left(-\frac{1}{3} - \frac{1}{9}S_{axis}^2\right) \frac{\tau'}{1 + (\omega\tau')^2} \quad [\text{S3}]$$

where τ_c is the rotational correlation time for the assumed spherical molecule to which the methyl group is attached, S_{axis}^2 is the generalized order parameter of the methyl 3-fold symmetry axis, $1/\tau' = 1/\tau_c + 1/\tau_e$, where τ_e is the time constant for rapid rotation of the methyl averaging axis. Assuming that $(\omega_C \tau_c)^2 \gg 1$, $(\omega_C \tau')^2 \ll 1$ it follows that

$$\Delta R_2 = 0.3d_{CH}^2 \left[\frac{S_{axis}^2}{\omega_C^2 \tau_c} + 3 \left(1 - \frac{S_{axis}^2}{9} \right) \tau' + 6 \left(-\frac{1}{3} - \frac{S_{axis}^2}{9} \right) \tau' \right] \approx 0.3d_{CH}^2 \left[\frac{S_{axis}^2}{\omega_C^2 \tau_c} + (1 - S_{axis}^2) \tau_e \right].$$

[S4]

Eq. [S4] can be compared with the corresponding expression for the ^{13}C longitudinal relaxation rate, $R_1(^{13}\text{C})$, for a $^{13}\text{CH}_3$ spin system (neglecting CSA) as measured using a pulse scheme that suppresses cross-correlation between ^{13}C – ^1H dipolar interactions^[23]

$$R_1(^{13}\text{C}) \approx 0.3d_{CH}^2 \left[\frac{3.35}{9} \frac{S_{axis}^2}{\omega_C^2 \tau_c} + 10 \left(1 - \frac{S_{axis}^2}{9} \right) \tau_e \right].$$

[S5]

It is noteworthy that although $J(0) \approx K(0)$ for intra-methyl ^{13}C – ^1H dipolar interactions, $J(\omega_C)$ is typically different from $K(\omega_C)$ due to the large contribution from τ_e ^[24], leading to differences between $R_1(^{13}\text{C})$ and ΔR_2 . For example, assuming $S_{axis}^2 = 0.5$, $\tau_e = 20$ ps and $\tau_c = 5$ ns, parameters that are similar to those for GB1 (25°C), the calculated $R_1(^{13}\text{C})/\Delta R_2$ ratio is 1.94 at 600 MHz, and the average measured value for GB1 at 600 MHz is 1.94, as well. Average values for ΔR_2 are 1.08 s⁻¹ for GB1 and 1.62 s⁻¹ for the FF domain (600 MHz).

Evolution of ^1H TQ coherences due to chemical exchange. In the simplest implementation the evolution of ^1H TQ coherences due to chemical shift, intra-methyl ^1H – ^{13}C scalar couplings, $^1J_{HC}$, and two-site chemical exchange, $G \xrightleftharpoons[k_{EG}]{k_{GE}} E$, is given by

$$\frac{d}{dt} \begin{pmatrix} M_A^G \\ M_I^G \\ M_A^E \\ M_I^E \end{pmatrix} = - \begin{pmatrix} 3i\omega_G + R_{2,A}^G + k_{GE} & 3i\pi^1J_{HC} & -k_{EG} & 0 \\ 3i\pi^1J_{HC} & 3i\omega_G + R_{2,I}^G + k_{GE} & 0 & -k_{EG} \\ -k_{GE} & 0 & 3i\omega_E + R_{2,A}^E + k_{EG} & 3i\pi^1J_{HC} \\ 0 & -k_{GE} & 3i\pi^1J_{HC} & 3i\omega_E + R_{2,I}^E + k_{EG} \end{pmatrix} \begin{pmatrix} M_A^G \\ M_I^G \\ M_A^E \\ M_I^E \end{pmatrix}$$

[S6]

where $M_A = 3C_z I_+^1 I_+^2 I_+^3$, $M_I = 1.5I_+^1 I_+^2 I_+^3$, the subscripts or superscripts G and E denote ground and excited states, respectively, and $R_{2,A}$ and $R_{2,I}$ are intrinsic transverse relaxation rates for TQ anti- and in-phase coherences. A similar set of equations holds for the evolution of $M_A = 3C_z I_-^1 I_-^2 I_-^3$, $M_I = 1.5I_-^1 I_-^2 I_-^3$ as well, with i replaced by $-i$. All analyses assume equal relaxation rates in ground and excited states. It should be noted that Eq. [S6] is a simplification, in general, as the number of basis set elements that are needed to account for offset effects, B_1 inhomogeneity and pulse miscalibration must be larger. Since TQ coherences can only be created within the subspace of the $I = 3/2$ manifold and pulses do not couple coherences belonging to different manifolds^[25] a more rigorous treatment would be to include a pair of 32 basis set elements that span the $3/2$ manifold. Each basis set includes both in-phase and anti-phase elements to account for scalar coupled ($^1J_{HC}$) evolution, one set for spins in the ground state and a second for spins in the excited state. We have found that the simple approach of Eq. [S6] produces essentially identical results to those obtained with the more complex treatment and have therefore used Eq. [S6] in all analyses of dispersion data.

As described above the fitting parameters include global values (k_{GE} , k_{EG}) as well as per-residue rates, $R_{2,A}$ and $\Delta R_2 = R_{2,I} - R_{2,A}$. In the case where flat dispersions are obtained, such as for GB1 (Figure 1), these are ‘corrected’ for differential relaxation between in-phase and anti-phase TQ coherences using the relation^[26]

$$R_{2,eff}(v_{CPMG}, \text{corrected}) = R_{2,eff}(v_{CPMG}) - \Delta R_2 \left[\frac{1}{2} - \frac{\sin(6\pi^1 J_{HC} \tau_{cp})}{12\pi^1 J_{HC} \tau_{cp}} \right] \quad [S7]$$

and no attempt at fitting the data has been made. By contrast, ΔR_2 is used as an explicit fitting parameter when dispersion profiles are fitted.

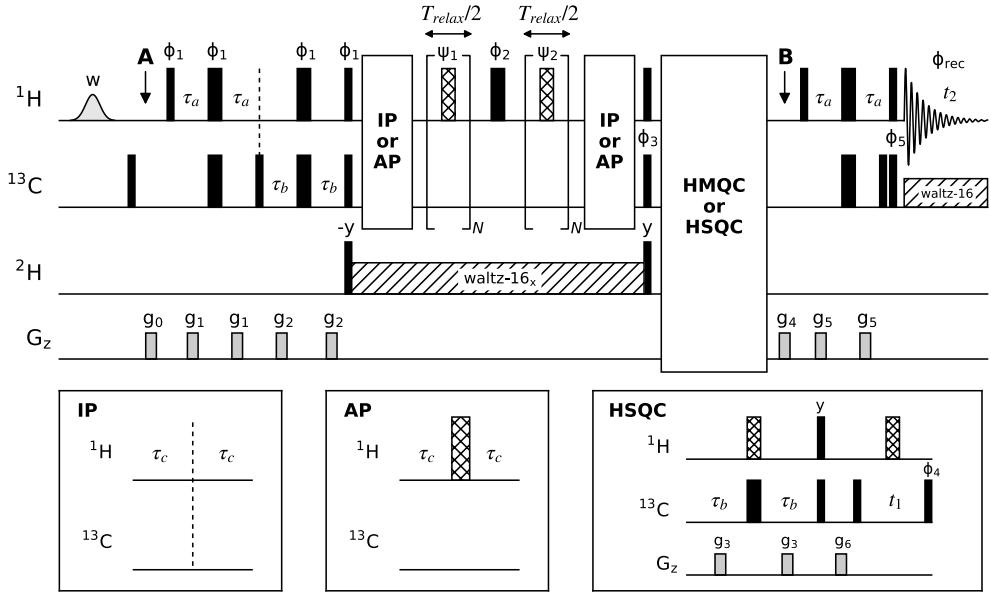


Figure S1. Pulse scheme for measurement of ^1H TQ CPMG relaxation dispersion profiles (see Figure 1) with ^{13}C chemical shifts recorded via either HMQC (Figure 1) or HSQC elements. Details are as per Figure 1, with $\tau_c = 1/(12^1J_{HC})$ ($\simeq 0.65$ ms) and g_6 : (0.5, -15). Insets labeled IP and AP are schemes for measuring the difference between transverse relaxation rates of anti-phase (AP) $3C_zI_+^1I_-^2I_+^3 + 3C_zI_-^1I_+^2I_-^3$, and in-phase (IP) $-1.5iI_+^1I_-^2I_+^3 + 1.5iI_-^1I_+^2I_-^3$, TQ elements. By recording a pair of planes (one for in-phase and one for anti-phase) at high ^1H pulsing rates ($v_{CPMG} = 2$ kHz), so that the interconversion between in-phase and anti-phase magnetization is prevented, the difference in R_2 rates is obtained from the relation $\Delta R_2 = R_{2,I} - R_{2,A} = -\frac{1}{T_{relax}} \ln \frac{I_I}{I_A}$ where I_I and I_A are intensities of corresponding peaks in spectra recording the evolution of in-phase and anti-phase coherences, respectively. Note that the IP and AP elements are removed when CPMG dispersion profiles are measured.

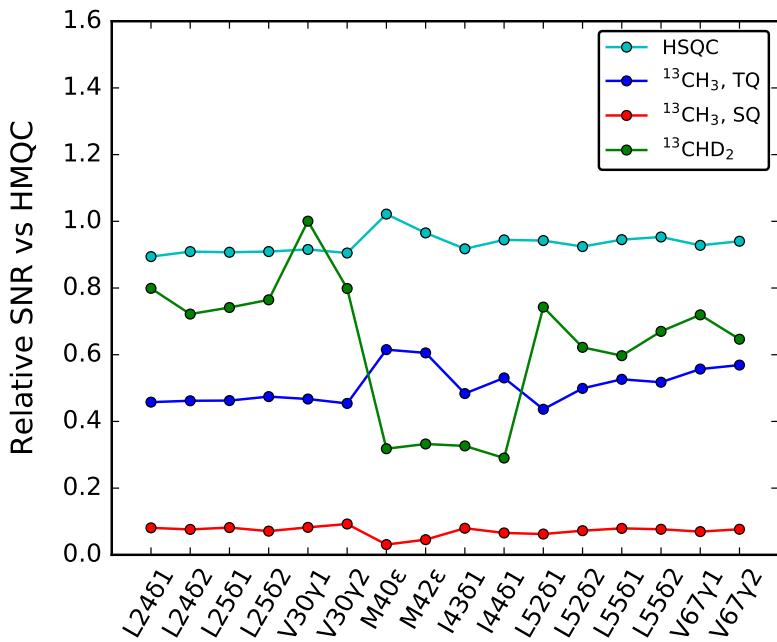


Figure S2. Per-residue signal to noise ratios (SNRs) as measured from $^{13}\text{C}-^1\text{H}$ HSQC (cyan), ^1H TQ CPMG RD (blue), ^1H SQ $^{13}\text{CHD}_2$ RD (green) and ^1H SQ $^{13}\text{CH}_3$ RD (red) schemes, recorded on ILVM- $^{13}\text{CH}_3$ or ILVM- $^{13}\text{CHD}_2$ FF domain samples, 600 MHz, 25°C. Intensities of correlations were measured in 2D spectra recorded with the CPMG element removed. All intensities are normalized to those obtained in a $^{13}\text{C}-^1\text{H}$ HMQC reference plane. Average SNRs are 0.93 ± 0.03 , 0.73 ± 0.10 (0.32 ± 0.02), 0.51 ± 0.05 and 0.07 ± 0.01 for HSQC, L,V residues(I,M residues) in $^{13}\text{CHD}_2$ SQ CPMG, $^{13}\text{CH}_3$ TQ CPMG, and $^{13}\text{CH}_3$ SQ CPMG, respectively. These values are in reasonably good agreement with calculations. For example, the relative SNR of L,V ($^{13}\text{CH}_3$) cross peaks in the TQ vs SQ ($^{13}\text{CHD}_2$) data sets is predicted to be $\frac{3 \times 0.75}{2\sqrt{2}} = 0.80$ (measured value of 0.68) where the factor of 3 takes into account that magnetization from 3 methyl protons contributes to the TQ signal, while 0.75 includes the loss from generation of TQ coherences. The factor of $2\sqrt{2}$ in the denominator reflects the fact that the effective concentration of NMR active L,V methyl groups is twice as high with the $^{13}\text{CHD}_2$ labeling scheme relative to $^{13}\text{CH}_3$ (both isopropyl methyls are $^{13}\text{CHD}_2$, while only one is $^{13}\text{CH}_3$) and an enhanced sensitivity pulse scheme is used that preserves both cosine and sine modulated t_1 components (factor of $\sqrt{2}$). In the case of I,M methyl groups the SNR in TQ and $^{13}\text{CHD}_2$ SQ data sets is predicted to be $\frac{3 \times 0.75}{\sqrt{2}} = 1.59$ and a value of 1.77 is measured.

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```

tsatpwr,      /* low level 1H trans.power for presat */
sw1,          /* sweep width in f1 */

tpwr_cp,      /* power level for 1H CPMG      */
pw_cp,         /* 1H pw for CPMG      */

ncyc_cp,      /* number of CPMG cycles */
time_T2,       /* total time for CPMG trains  */
tau_cpmg,     /*

tof_me,       /*

dhpwr,        /*

pwc,          /*

dpwr3,        /*

dpwr3_D,      /*

pwd,          /*

pwd1,          /*

gt0,          /*

gt1,          /*

gt2,          /*

gt3,          /*

gt4,          /*

gt5,          /*

gt6,          /*

gzlvl10,      /*

gzlvl11,      /*

gzlvl12,      /*

gzlvl13,      /*

gzlvl14,      /*

gzlvl15,      /*

gzlvl16,      /*

tpwrs1,       /*

pw_s1;        /*

/* LOAD VARIABLES */

getstr("fsat",fsat);
getstr("f1180",f1180);
getstr("fscuba",fscuba);

getstr("shape_s1",shape_s1);
getstr("water_flg",water_flg);
getstr("mq_flg",mq_flg);
getstr("cal_flg",cal_flg);
getstr("AP_flg",AP_flg);
getstr("D_flg",D_flg);
getstr("comp180_flg",comp180_flg);

tau_a = getval("tau_a");
tau_b = getval("tau_b");
tau_c = getval("tau_c");
tpwr = getval("tpwr");
tsatpwr = getval("tsatpwr");
dpwr = getval("dpwr");
phase = (int) (getval("phase") + 0.5);
sw1 = getval("sw1");

tpwr_cp = getval("tpwr_cp");
pw_cp = getval("pw_cp");
ncyc_cp = getval("ncyc_cp");
time_T2 = getval("time_T2");

tof_me = getval("tof_me");

dpwr3 = getval("dpwr3");
dpwr3_D = getval("dpwr3_D");

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pwd = getval("pwd");
pwd1 = getval("pwd1");

dhpwr = getval("dhpwr");
pwc = getval("pwc");

gt0 = getval("gt0");
gt1 = getval("gt1");
gt2 = getval("gt2");
gt3 = getval("gt3");
gt4 = getval("gt4");
gt5 = getval("gt5");
gt6 = getval("gt6");

gzlvl0 = getval("gzlvl0");
gzlvl1 = getval("gzlvl1");
gzlvl2 = getval("gzlvl2");
gzlvl3 = getval("gzlvl3");
gzlvl4 = getval("gzlvl4");
gzlvl5 = getval("gzlvl5");
gzlvl6 = getval("gzlvl6");

tpwrsl = getval("tpwrsl");
pw_sl = getval("pw_sl");

/* LOAD PHASE TABLE */

settable(t1,6,phi1);
settable(t2,2,phi2);
settable(t3,12,phi3);
settable(t4,24,phi4);
settable(t5,2,phi5);
settable(t11,16,psi1);
settable(t12,16,psi2);
settable(t13,16,psi3);
settable(t14,16,psi4);
settable(t15,16,psi5);
settable(t16,16,psi5);
settable(t8,24,rec);

/* CHECK VALIDITY OF PARAMETER RANGES */

if((dm[A] == 'y' || dm[B] == 'y' || dm[C] == 'y'))
{
    printf("incorrect dec1 decoupler flags!  ");
    abort();
}

if((dm2[A] == 'y' || dm2[B] == 'y' || dm2[C] == 'y' || dm2[D] == 'y'))
{
    printf("incorrect dec2 decoupler flags!  ");
    abort();
}

if( tsatpwr > 6 )
{
    printf("TSATPWR too large !!!  ");
    abort();
}

if( pw > 200.0e-6 )
{
    printf("dont fry the probe, pw too high !  ");
    abort();
}

if( tpwr_cp > 61 )
{
    printf("don't fry the probe, tpwr_cp too large: < 62!  ");
    abort();
}

```

```

if( pw_cp < 9.0e-6 ) {
    printf("pw_cp is too low; > 9.0us\n");
    abort();
}

if( dpwr > 48 )
{
    printf("don't fry the probe, DPWR too large!  ");
    abort();
}

if( dpwr2 > -16 )
{
    printf("don't fry the probe, DPWR2 too large!  ");
    abort();
}

if( gt0 > 3e-3 || gt1 > 3e-3 || gt2 > 3e-3 || gt3 > 3e-3 || gt4 > 3e-3 || gt5 >
3e-3
            || gt6 > 3e-3 )
{
    printf("gradients on for too long. Must be < 3e-3 \n");
    abort();
}

if( ncyc_cp > 80 ) {
    printf("ncyc_cp is too large; must be less than 81\n");
    abort();
}

if( time_T2 > 0.080 ) {
    printf("time_T2 is too large; must be less than 80 ms\n");
    abort();
}

if( ncyc_cp > 0 ) {
    tau_cpmg = time_T2/(4.0*ncyc_cp);
    if(ix == 1)
        printf("nuCPMG for current experiment is (Hz): %5.3f\n",1/(4.0*tau_cpmg));
} else {
    tau_cpmg = time_T2/(4.0);
    if(ix == 1)
        printf("minimum nuCPMG for current experiment is (Hz): %5.3f\n",1./(4.0*tau
_cpmg));
}

if( tau_cpmg < 50e-6 ) {
    printf("tau_cpmg is too small; decrease ncyc_cp\n");
    abort();
}

if( dpwr3 > 59 ) {
    printf("dpwr3 is too high; < 60\n");
    abort();
}

if( dpwr3_D > 51 ) {
    printf("dpwr3 is too high; < 52\n");
    abort();
}

/* Phase incrementation for hypercomplex 2D data */

if (phase == 2){
    tsadd(t4,1,4);
}

/* Set up f1180  tau1 = t1
   tau1 = d2;

if(f1180[A] == 'y') {
    if (mq_flg[A] == 'y') {

```

```

        tau1 += ( 1.0 / (2.0*sw1) ) - 4.0*pw - 4.0e-6;
    }else{
        tau1 += ( 1.0 / (2.0*sw1) ) - 4.0*pwc/PI - 4.0*pw - 4.0e-6;
    }
    if(tau1 < 0.4e-6) tau1 = 0.4e-6;
}

if(tau1 < 0.4e-6)
    tau1 = 0.4e-6;

tau1 = tau1/2.0;

/* Calculate modifications to phases for States-TPPI acquisition */

if(ix == 1) d2_init = d2 ;
t1_counter = (int) ( (d2-d2_init)*sw1 + 0.5 );
if(t1_counter % 2) {
    tsadd(t4,2,4);
    tsadd(t8,2,4);
}

/* BEGIN ACTUAL PULSE SEQUENCE */

status(A);

rlpower(tsatpwr,TODEV);      /* Set transmitter power for 1H presaturation */
rlpower(dhpwr,DODEV);        /* Set Decl power for 13C CPMG pulses */
rlpower(dpwr2,DO2DEV);        /* Set Dec2 power for 15N decoupling */

obsoffset(tof);

/* Presaturation Period */

status(B);

lk_sample();

if (fsat[0] == 'y')
{
    delay(2.0e-5);
    rgpulse(d1,zero,2.0e-6,2.0e-6); /* presaturation */
    rlpower(tpwr,TODEV); /* Set transmitter power for hard 1H pulses */
    delay(2.0e-5);
    if(fscuba[0] == 'y')
    {
        delay(2.2e-2);
        rgpulse(pw,zero,2.0e-6,0.0);
        rgpulse(2*pw,one,2.0e-6,0.0);
        rgpulse(pw,zero,2.0e-6,0.0);
        delay(2.2e-2);
    }
}
else
{
    delay(d1);
}

rlpower(tpwr_cp,TODEV);        /* Set transmitter power for 1H CPMG pulses */
txphase(zero);
dec2phase(zero);
decphase(zero);
delay(1.0e-5);

/* Begin Pulses */

status(C);

rcvroff();
lk_hold();
delay(20.0e-6);

if(water_flg[A] == 'y') {
    obspower(tpwrs1);
}

```

```

    shaped_pulse(shape_sl,pw_sl,zero,4.0e-6,0.0);
    obspower(tpwr_cp);
}

decrgpulse(pwc,zero,4.0e-6,0.0);

obsoffset(tof_me);
obsstepsize(60.0);
xmtrphase(t1);

delay(2.0e-6);
rgradient('z',g兹l10);
delay(gt0);
rgradient('z',0.0);
delay(250.0e-6);

rgpulse(pw_cp,zero,0.0,0.0);

delay(2.0e-6);
rgradient('z',g兹l11);
delay(gt1);
rgradient('z',0.0);
delay(250.0e-6);

delay(taua - gt1 - 252.0e-6 - (2.0/PI)*pw_cp);
simpulse(2.0*pw_cp,2.0*pwc,zero,zero,0.0,0.0);

delay(2.0e-6);
rgradient('z',g兹l11);
delay(gt1);
rgradient('z',0.0);
delay(250.0e-6);

delay(taua - gt1 - 252.0e-6);

decrgpulse(pwc,zero,0.0,0.0);

delay(2.0e-6);
rgradient('z',g兹l12);
delay(gt2);
rgradient('z',0.0);
delay(250.0e-6);

delay(taub - gt2 - 252.0e-6);
simpulse(2.0*pw_cp,2.0*pwc,zero,zero,0.0,0.0);

delay(taub - gt2 - 252.0e-6 - POWER_DELAY - pwd1 - 4.0e-6 - POWER_DELAY - PRG_START_DELAY);

delay(2.0e-6);
rgradient('z',g兹l12);
delay(gt2);
rgradient('z',0.0);
delay(250.0e-6);

if (D_flg[A] == 'y') {
    dec3power(dpwr3);
    dec3rgpulse(pwd1,three,4.0e-6,0.0);
    dec3phase(zero);
    dec3power(dpwr3_D);
    dec3unblank();
    dec3prgon(dseq3,pwd,dres3);
    dec3on();
} else {
    delay(POWER_DELAY + pwd1 + 4.0e-6 + POWER_DELAY + PRG_START_DELAY);
}

decrgpulse(pwc,zero,0.0,0.0);
rgpulse(pw_cp,zero,0.0,4.0e-6);

xmtrphase(zero);

```

```

decphase(t3);

if (cal_flg[A] == 'y') {
    if (AP_flg[A] == 'y') {
        delay(tauc - 2.0*pw_cp - WFG_START_DELAY - 4.0e-6);
        shaped_pulse("composite_1", 4.0*pw_cp, zero, 4.0e-6, 0.0);
        delay(tauc - 2.0*pw_cp - WFG_STOP_DELAY);
    }
    else {
        delay(2.0*tauc);
    }
}

if (ncyc_cp == 1) {
    delay(tau_cpmg - 2.0*pw_cp - WFG_START_DELAY - 4.0e-6);
    shaped_pulse("composite_1", 4.0*pw_cp, zero, 4.0e-6, 0.0);
    delay(tau_cpmg - 2.0*pw_cp - WFG_STOP_DELAY);
}

if (ncyc_cp > 1) {
    assign(zero,v1);

    initval(ncyc_cp,v4);
    loop(v4,v5);
    if (comp180_flg[A] == 'y') {
        getelem(t11,v1,v2);
        delay(tau_cpmg - 2.0*pw_cp - WFG_START_DELAY - 4.0e-6);
        shaped_pulse("composite_1", 4.0*pw_cp, v2, 4.0e-6, 0.0);
        delay(tau_cpmg - 2.0*pw_cp - WFG_STOP_DELAY);
    }
    else {
        if((int) ncyc_cp % 8 == 3 || (int) ncyc_cp % 8 == 5) {
            getelem(t11,v1,v2);
        } else if((int) ncyc_cp % 8 == 1 || (int) ncyc_cp % 8 == 7) {
            getelem(t13,v1,v2);
        } else {
            getelem(t15,v1,v2);
        }
        delay(tau_cpmg - pw_cp - 4.0e-6);
        rgpulse(2.0*pw_cp,v2,4.0e-6,0.0);
        delay(tau_cpmg - pw_cp);
    }
    incr(v1);
    endloop(v5);
}

rgpulse(2.0*pw_cp,t2,4.0e-6,4.0e-6);

if (ncyc_cp == 1) {
    delay(tau_cpmg - 2.0*pw_cp - WFG_START_DELAY - 4.0e-6);
    shaped_pulse("composite_2", 4.0*pw_cp, zero, 4.0e-6, 0.0);
    delay(tau_cpmg - 2.0*pw_cp - WFG_STOP_DELAY);
}

if (ncyc_cp > 1) {
    decr(v1);

    initval(ncyc_cp,v4);
    loop(v4,v5);
    if (comp180_flg[A] == 'y') {
        getelem(t12,v1,v2);
        delay(tau_cpmg - 2.0*pw_cp - WFG_START_DELAY - 4.0e-6);
        shaped_pulse("composite_2", 4.0*pw_cp, v2, 4.0e-6, 0.0);
        delay(tau_cpmg - 2.0*pw_cp - WFG_STOP_DELAY);
    }
    else {
        if((int) ncyc_cp % 8 == 3 || (int) ncyc_cp % 8 == 5) {
            getelem(t12,v1,v2);
        } else if((int) ncyc_cp % 8 == 1 || (int) ncyc_cp % 8 == 7) {
            getelem(t14,v1,v2);
        } else {
            getelem(t16,v1,v2);
        }
        delay(tau_cpmg - pw_cp - 4.0e-6);
        rgpulse(2.0*pw_cp,v2,4.0e-6,0.0);
    }
}

```

```

        delay(tau_cpmg - pw_cp);
    }
    decr(v1);
endloop(v5);
}

if (cal_flg[A] == 'y') {
    if (AP_flg[A] == 'y') {
        delay(tauc - 2.0*pw_cp - WFG_START_DELAY - 4.0e-6);
        shaped_pulse("composite_2", 4.0*pw_cp, zero, 4.0e-6, 0.0);
        delay(tauc - 2.0*pw_cp - WFG_STOP_DELAY);
    }
    else{
        delay(2.0*tauc);
    }
}

rgpulse(pw_cp, zero, 4.0e-6 + SAPS_DELAY, 0.0);
decrgpulse(pwc, t3, 0.0, 0.0);

if (D_flg[A] == 'y') {
    dec3off();
    dec3prgoff();
    dec3blank();
    dec3power(dpwr3);
    dec3rgpulse(pwd1, one, 4.0e-6, 0.0);
} else{
    delay(POWER_DELAY + PRG_STOP_DELAY + pwd1 + 4.0e-6);
}

delay(2.0e-6);
rgradient('z', gzlvl3);
delay(gt3);
rgradient('z', 0.0);
delay(250.0e-6);

decphase(zero);

if (mq_flg[A] == 'y') {
    delay(taub - gt3 - 252.0e-6 + WFG_START_DELAY - POWER_DELAY - PRG_STOP_DELAY
- pwd1 - 4.0e-6);

    decrgpulse(2.0*pwc, zero, 0.0, 0.0);

    delay(taul);
    shaped_pulse("composite_1", 4.0*pw_cp, zero, 4.0e-6, 0.0);
    delay(taul);

    txphase(one);
    decphase(t4);

    delay(2.0e-6);
    rgradient('z', gzlvl3);
    delay(gt3);
    rgradient('z', 0.0);
    delay(250.0e-6);

    delay(taub - gt3 - 252.0e-6 - WFG_STOP_DELAY);

    decrgpulse(pwc, t4, 0.0, 0.0);

    delay(2.0*pwc + 2.0*WFG_START_DELAY);

    rgpulse(pw_cp, one, 0.0, 0.0);

} else{
    delay(taub - gt3 - 252.0e-6 - WFG2_START_DELAY - POWER_DELAY - PRG_STOP_DELAY
- pwd1 - 8.0e-6);

    simshaped_pulse("composite_1", "hard", 4.0*pw_cp, 2.0*pwc, zero, zero, 4.0e-6, 0.0);
    txphase(one);
}

```

```

        delay(2.0e-6);
        rgradient('z',gzlvl3);
        delay(gt3);
        rgradient('z',0.0);
        delay(250.0e-6);

        delay(taub - gt3 - 252.0e-6 - WFG2_STOP_DELAY);

        decrgpulse(pwc,zero,0.0,0.0);
        rgpulse(pw_cp,one,0.0,0.0);

        delay(2.0e-6);
        rgradient('z',gzlvl6);
        delay(gt6);
        rgradient('z',0.0);
        delay(250.0e-6);

        decrgpulse(pwc,zero,0.0,0.0);
        decphase(t4);

        delay(tau1);

        shaped_pulse("composite_1",4.0*pw_cp,zero,4.0e-6,0.0);

        delay(tau1);

        decrgpulse(pwc,t4,0.0,0.0);
    }

    obsoffset(tof);
    decphase(zero);
    obspower(tpwr);

    delay(2.0e-6);
    rgradient('z',gzlvl4);
    delay(gt4);
    rgradient('z',0.0);
    delay(250.0e-6);

    rgpulse(pw,zero,2.0e-6,0.0);

    delay(2.0e-6);
    rgradient('z',gzlvl5);
    delay(gt5);
    rgradient('z',0.0);
    delay(150.0e-6);

    delay(taua - gt5 - 152.0e-6);

    simpulse(2.0*pw,2.0*pwc,zero,zero,0.0,0.0);

    delay(2.0e-6);
    rgradient('z',gzlvl5);
    delay(gt5);
    rgradient('z',0.0);
    delay(150.0e-6);

    delay(taua - gt5 - 152.0e-6 - pwc - 2.0e-6 - pwc - 2.0*POWER_DELAY);

    decrgpulse(pwc,zero,0.0,0.0);
    decrgpulse(pwc,t5,2.0e-6,0.0);

    rlpower(dpwr,DODEV); /* Set power for decoupling */
    rlpower(dpwr2,DO2DEV); /* Set power for decoupling */

    rgpulse(pw,zero,0.0,0.0);

/* BEGIN ACQUISITION */

status(D);
    setreceiver(t8);
}

```