

Supporting Information for Yang and Kay, " TROSY Triple Resonance Four Dimensional Spectroscopy of a 46 ns Tumbling Protein"

Figure 1: TROSY-based triple resonance pulse schemes, CT-HNCA, CT-HN(CO)CA, CT-HN(CA)CB, CT-HN(COCA)CB, which make use of the enhanced sensitivity pulsed field gradient implementation for the transfer of magnetization from  $^{15}\text{N}$  to HN prior to detection<sup>1</sup>. The shaded region of each of sequences A - D can be replaced with either schemes (a) or (b). In scheme (a)  $\phi_0 = x$  and suppression of the undesired crosspeak derived from the pathway,  $\text{N}_{\text{TR}}(1 + 2\text{HN}_Z) \rightarrow \text{HN}_{\text{TR}}(1 - 2\text{N}_Z)$ , is achieved by efficient relaxation occurring during the  $\tau_b$  and  $\text{TN}$  delays (totaling  $\approx 50$  ms) and the broad  $^1\text{H}$  linewidth<sup>1</sup>. In sequence (b) active suppression of the undesired crosspeak occurs by actively selecting only the slowly relaxing component<sup>2</sup>. The value of  $\tau_e$  is set to  $1/(8J_{\text{NH}})$ , while  $|\phi_0| = 45^\circ$ . The sign of  $\phi_0$  must be adjusted on each brand of spectrometer to give optimum sensitivity. Note that the phase of the final  $^1\text{H}$   $90^\circ$  of the INEPT<sup>3</sup> ( $y$  or  $-y$ ) must also be adjusted for optimum signal and can vary between spectrometers. (The phase values listed were calibrated on a Varian spectrometer). The  $^1\text{H}$   $180^\circ$  pulse applied after a delay  $\tau_e$  from the start of  $^{15}\text{N}$  evolution is of the composite variety ( $90_x 180_y 90_x$ ). The shaped  $^1\text{H}$   $90^\circ$  pulse is a water selective pulse having the EBURP-1 profile<sup>4</sup> (1.05 kHz). Use of scheme (b) does result in a slight attenuation of the signal relative to sequence (a) since for the first  $\tau_e$  delay the fast relaxing  $^{15}\text{N}$  component is the one which ultimately contributes to the signal of interest. Nevertheless a comparison of TROSY based triple resonance experiments recorded with scheme (b) using the enhanced sensitivity pulsed field gradient approach<sup>1</sup> was 3% more sensitive than the implementation of Pervushin et al.<sup>5</sup> for an MBP sample (600 MHz) at  $37^\circ\text{C}$  ( $\tau_C = 17$  ns) and at  $25^\circ\text{C}$  ( $\tau_C = 23$  ns). It is difficult to provide a definitive statement regarding when to use scheme (a) or (b). Figure 1 of Yang and Kay<sup>1</sup> illustrates the relative intensity ratio of the crosspeaks derived from the two pathways [  $\text{N}_{\text{TR}}(1 \pm 2\text{HN}_Z) \rightarrow \text{HN}_{\text{TR}}(1 \mp 2\text{N}_Z)$  ] which can be used as a guideline. For

spectra recorded on MBP at 25 °C ( $\tau_C = 23$  ns) using scheme (a) the intensity of the undesired component was no more than 5-6% of the slowly relaxing crosspeak and is predicted to be under 2% for spectra recorded at 800 MHz<sup>1</sup>. The undesired crosspeak is below the noise level in spectra recorded on MBP at 5 °C and at 600 MHz. In order to establish whether scheme (a) or (b) should be employed a 2D <sup>15</sup>N-<sup>1</sup>HN plane of an HNCO can be recorded to evaluate the size of the undesired crosspeak when sequence a is employed. In what follows only a brief description of the pulse schemes is provided; details can be found in the original references<sup>6-8</sup>. A. CT-TROSY-HNCA. The <sup>13</sup>C carrier is centered at 58 ppm and the rectangular <sup>13</sup>C<sup>α</sup> 90° (<sup>13</sup>C<sup>α</sup>/<sup>13</sup>C' 180°) pulses are applied with a field strength of  $\Delta/\sqrt{15}$  ( $\Delta/\sqrt{3}$ )<sup>9</sup>, where  $\Delta$  is defined as in the legend to Figure 1. The shaped non-selective <sup>13</sup>C<sup>α</sup> refocusing pulse has the REBURP<sup>4</sup> profile and is centered at 45 ppm. The phase cycle used is:  $\phi_1 = 2(x), 2(-x)$ ;  $\phi_2 = (y, -y)$ ;  $\phi_3 = 4(x), 4(-x)$ ;  $\phi_4 = x$ ;  $\text{rec} = x, 2(-x), x$ . B. CT-TROSY-HN(CO)CA. The <sup>13</sup>C carrier is initially set to 176 ppm, jumped to 58 ppm prior to the application of the <sup>13</sup>C<sup>α</sup> 90° pulse of phase  $\phi_2$  and set to 176 ppm after the subsequent <sup>13</sup>C<sup>α</sup> 90° pulse. All <sup>13</sup>C' and <sup>13</sup>C<sup>α</sup> 90° pulses are applied with a field of  $\Delta/\sqrt{15}$ ; the <sup>13</sup>C<sup>α</sup> and <sup>13</sup>C' rectangular 180° pulses have a strength of  $\Delta/\sqrt{3}$ ; the <sup>13</sup>C<sup>α</sup> shaped pulse during the CT-<sup>13</sup>C<sup>α</sup> evolution period has the REBURP profile<sup>4</sup> with the center of excitation at 45 ppm. Vertical arrows indicate the position of the Bloch-Siegert compensation pulses<sup>10</sup>. Note that the <sup>13</sup>C<sup>α</sup> 180° pulse during the first (second)  $2\tau_C$  period is applied after (before) the <sup>13</sup>C' pulse. The phase cycle is:  $\phi_1 = (x, -x)$ ;  $\phi_2 = 2(x), 2(-x)$ ;  $\phi_3 = y$ ;  $\phi_4 = 4(x), 4(-x)$ ;  $\phi_5 = x$ ;  $\text{rec} = x, 2(-x), x$ . C. CT-TROSY-HN(CA)CB. The <sup>13</sup>C carrier is at 43 ppm and the <sup>13</sup>C<sup>α</sup> pulse (of schemes a and b) and the final <sup>13</sup>C<sup>α/β</sup> 180° pulse labeled  $\alpha$  are applied with the center of excitation at 58 ppm by phase modulation of the carrier<sup>11,12</sup>. All <sup>13</sup>C<sup>α/β</sup> 90° pulses are applied at the highest field possible, while the shaped (non-selective for aliphatic carbons) <sup>13</sup>C<sup>α/β</sup> refocusing pulses have the REBURP profile<sup>4</sup>. The phases of these pulses are carefully adjusted for optimum sensitivity. The <sup>13</sup>C' pulse labeled with \* is applied with the G3 profile<sup>13</sup> ( $\approx 300$   $\mu$ s at 600 MHz) centered

at 150 ppm. The phase cycle employed is:  $\phi_0 = (x,-x)$  for scheme (a) and either (45°, 225°) or (-45°, 135°) for scheme (b), depending on the brand of spectrometer;  $\phi_1 = 2(x), 2(-x)$ ;  $\phi_2 = 4(y), 4(-y)$ ;  $\phi_3 = y$ ;  $\phi_4 = 4(x), 4(-x)$ ;  $\phi_5 = x$ ;  $\text{rec} = x, 2(-x), x$ . D. CT-TROSY-HN(COCA)CB. The  $^{13}\text{C}$  carrier is initially set to 176 ppm, jumped to 43 ppm prior to the application of the  $^{13}\text{C}^{\alpha/\beta}$  90° pulse of phase  $\phi_2$  and set back to 176 ppm prior to the  $^{13}\text{C}'$  90<sub>y</sub> pulse. All  $^{13}\text{C}^{\alpha/\beta}$  90° pulses are applied with a field of  $\Delta'/\sqrt{15}$ ;  $^{13}\text{C}'$  90° pulses are applied with a field strength of  $\Delta/\sqrt{15}$ ; the  $^{13}\text{C}'$  rectangular 180° pulses have a strength of  $\Delta/\sqrt{3}$ ; the  $^{13}\text{C}^{\alpha/\beta}$  shaped pulses during the CT- $^{13}\text{C}^{\beta}$  evolution period and the two  $2x\tau_d$  periods have REBURP<sup>4</sup> profiles with the center of excitation at 43 ppm; the  $^{13}\text{C}$  pulses labeled  $\alpha$  [and the first  $^{13}\text{C}^{\alpha}$  pulse of either scheme (a) or (b)] are centered at 58 ppm by phase modulation of the carrier<sup>11,12</sup> and applied with a field of  $\Delta/\sqrt{3}$ . The value of  $\Delta'$  is set to the difference in Hz between the centers of the  $^{13}\text{C}'$  (176 ppm) and  $^{13}\text{C}^{\alpha/\beta}$  (43 ppm) spectral regions. Vertical arrows indicate the position of the Bloch-Siegert compensation pulses<sup>10</sup>. Other pulses are as defined above. The phase cycle is:  $\phi_1 = (x,-x)$ ;  $\phi_2 = 2(x), 2(-x)$ ;  $\phi_3 = 4(y), 4(-y)$ ;  $\phi_4 = y$ ;  $\phi_5 = 4(x), 4(-x)$ ;  $\phi_6 = x$ ;  $\text{rec} = x, 2(-x), x$ .

It is noteworthy that many of the original schemes used a  $^{13}\text{C}'$  180° pulse having the seduce-1 profile<sup>14</sup> rather than rectangular pulses with nulls at the center of the  $^{13}\text{C}^{\alpha}$  region. We have found these different pulses make little difference to the quality of the spectra. Note that on systems where a power change is accompanied by phase changes the phase adjustments needed may vary depending on whether the seduce profile or a rectangular pulse is employed.

## References

- (1) Yang, D.; Kay, L. E. *J. Biomol. NMR* **1999**, *13*, 3-9.
- (2) Pervushin, K.; Riek, R.; Wider, G.; Wüthrich, K. *J. Am. Chem. Soc.* **1998**, *120*, 6394-6400.

- (3) Morris, G. A.; Freeman, R. *J. Am. Chem. Soc.* **1979**, *101*, 760-762.
- (4) Geen, H.; Freeman, R. *J. Magn. Reson.* **1991**, *93*, 93-141.
- (5) Pervushin, K. V.; Wider, G.; Wüthrich, K. *J. Biomol. NMR* **1998**, *12*, 345-348.
- (6) Yamazaki, T.; Lee, W.; Revington, M.; Mattiello, D. L.; Dahlquist, F. W.; Arrowsmith, C. H.; Kay, L. E. *J. Am. Chem. Soc.* **1994**, *116*, 6464-6465.
- (7) Yamazaki, T.; Lee, W.; Arrowsmith, C. H.; Muhandiram, D. R.; Kay, L. E. *J. Am. Chem. Soc.* **1994**, *116*, 11655-11666.
- (8) Shan, X.; Gardner, K. H.; Muhandiram, D. R.; Rao, N. S.; Arrowsmith, C. H.; Kay, L. E. *J. Am. Chem. Soc.* **1996**, *118*, 6570-6579.
- (9) Kay, L. E.; Ikura, M.; Tschudin, R.; Bax, A. *J. Magn. Reson.* **1990**, *89*, 496-514.
- (10) Vuister, G. W.; Bax, A. *J. Magn. Reson.* **1992**, *98*, 428-435.
- (11) Patt, S. L. *J. Magn. Reson.* **1992**, *96*, 94-102.
- (12) Boyd, J.; Scoffo, N. *J. Magn. Reson.* **1989**, *85*, 406-413.
- (13) Emsley, L.; Bodenhausen, G. *Chem. Phys. Lett.* **1987**, *165*, 469-476.
- (14) McCoy, M.; Mueller, L. *J. Am. Chem. Soc.* **1992**, *114*, 2108-2110.



