

## CURRICULUM VITAE

### **Lewis Edward Kay** **OC, FRS, FRSC**

Department of Molecular Genetics,  
Department of Biochemistry  
Department of Chemistry  
University of Toronto  
Medical Sciences Building  
1 King's College Circle  
Toronto, Ontario, Canada, M5S 1A8

#### **Academic Degrees**

- 1988 PhD in Molecular Biophysics, Yale University, New Haven, Connecticut, USA.  
Supervisor: Dr. J.H. Prestegard; Thesis: "NMR Methods for Studying Motional and Structural Properties of Molecules in Solution"
- 1983 BSc in Biochemistry (first class honours), University of Alberta, Edmonton, Alberta, Canada

#### **Employment**

- 1995 - Professor of Molecular Genetics (previously Medical Genetics), Biochemistry and Chemistry, University of Toronto, Toronto, Ontario, Canada  
Phone: 416-978-0741, Fax: 416-978-8548, E-mail: kay@pound.med.utoronto.ca
- 1992 - 1995 Assistant Professor of Molecular Genetics (previously Medical Genetics), Biochemistry and Chemistry, University of Toronto, Toronto, Ontario, Canada
- 1988 - 1991 Alberta Heritage and Medical Research Council of Canada Post-Doctoral Fellow, Laboratory of Dr. Adrian Bax, Laboratory of Chemical Physics, National Institutes of Health, Bethesda, Maryland, USA

**Endowed Lectures (plenary lectures not listed here)**

Birss Lecturer, Department of Chemistry, University of Alberta, Edmonton, October 1999.

NIH Director's Wednesday Afternoon Lecture Series, May, 2006.

Wilbur Cross Medal Lecture, Yale University, Oct, 2006.

The Gitte Vold Memorial Lecture, University of California, San Diego, March 2007.

Proctor and Gamble Lecture, Department of Chemistry, UMass Amherst, October 2007

Hartek Lecturer, Department of Chemistry, RPI, October 2007

McDowell Lecture, Department of Chemistry, UBC, October 2009.

Colter Lecture, Department of Biochemistry, University of Alberta, Fall 2011.

Aline U. and James M. Orten Memorial Lecturer, Department of Biochemistry and Molecular Biology, Wayne State University, April 2012.

Sackler Lecturer, Tel Aviv University, 2013-2014.

Sir Rex Richards Lecture, Dept of Chemistry, Oxford, April 2016.

National Lecturer, Biophysical Society of Canada, Winnipeg, Manitoba, June, 2016.

2016 Distinguished Lecturer, Scripps Research Institute, La Jolla, CA, Sept. 2016.

Peng Memorial Lecture, Biophysics, University of Connecticut Health Center, Oct. 2016.

Robert Olson Lecture, Washington University & University of St. Louis, Dec, 2017.

2018 Lemieux Lecture, Dept. of Chemistry, University of Ottawa, May 2018.

Krimm Lecture, Dept. of Biophysics, University of Michigan, April 2019.

John T. Edsall Lecture, Molecular and Cellular Biology, Harvard University, May 2019.

2019 Garner King Lecture, Faculty of Medicine, University of Alberta, June 2019.

Maude Menten Lecture, Western University, October 2019. (Postponed)

Silvestri Lecture, Villanova, December 2020.

Nordin Lecture, U. Mass, Amherst, May 2023.

## Honors and Awards

- 2023 Akutsu Prize, Korean Magnetic Resonance Society, Seoul, Korea
- 2023 Honorary Doctorate of Science, University of Ottawa
- 2020 Ranked #2 in biophysics category of 2020 Stanford List of the world's top scientists
- 2020 Elected International Member of the National Academy of Sciences, USA
- 2019 Honorary Doctorate of Science, University of British Columbia
- 2018 Nakanishi Prize, American Chemical Society
- 2018 Gerhard Herzberg Canada Gold Medal for Science and Engineering
- 2017 Protein Society Christian B Anfinsen Award
- 2017 Canada Gairdner Foundation International Award
- 2017 Fellow of the Biophysical Society Award
- 2016 Appointed Officer of the Order of Canada
- 2016 Fellow of the Biophysical Society of Canada
- 2013 E. W. R. Steacie Award, Canadian Society for Chemistry
- 2012 University Professor, University of Toronto
- 2012 Khorana Prize, Royal Society of Chemistry, UK
- 2010 Election to the Royal Society of London
- 2009 Distinguished Alumni Award, University of Alberta
- 2008 Premier's Discovery Award, Province of Ontario
- 2008 Elected as one of the initial fellows of the International Society of Magnetic Resonance
- 2007 Dales Prize, Faculty of Medicine, University of Toronto
- 2006 Election to the Royal Society of Canada

2006 Wilbur Cross Medal of the Yale University Graduate School.

2005 Listed in ISI's database of Highly Cited Researchers; top 0.5% of most cited chemists in the world.

2004 Gunther Laukien Prize, Experimental NMR Conference.

2002 Founders Medal from the International Society of Magnetic Resonance in Biological Systems

2002 Flavelle Medal from the Royal Society of Canada

2000 Canada Research Chair, Tier 1

1999 Premier's Research Excellence Award, Province of Ontario

1999 Steacie Prize from the National Research Council of Canada

1999 Medical Research Council of Canada Scientist Award (1999-2000)

1998 University of Toronto McLean Award

1998 Canada's Top 40 Under 40 Award

1997 Member Editorial Board of Journal of Biomolecular NMR (- current)

1997 International Research Scholar of the Howard Hughes Medical Institute (1997-2002)

1996 Member Editorial Advisory Board of Biochemistry (-current)

1996 Merck Frosst Award

1995 Alfred P. Sloan Research Fellow (1995-1997)

1994 - 1999 MRC (Medical Research Council of Canada) Scholarship

1994 Editorial Board of Concepts in Magnetic Resonance (1994-1999)

1994 Appointed to the Faculty of NMR Concepts, Rhode Island, USA (1994-1999)

1993 - Editorial Board of the Journal of Magnetic Resonance (-current)

1990 - 1992 MRC (Medical Research Council of Canada) Centennial Fellowship

1988 - 1991 Alberta Heritage Post-Doctoral Fellowship

- 1988 - 1989 MRC (Medical Research Council of Canada) Post-Doctoral Fellowship
- 1983 - 1987 NSERC (Natural Science and Engineering Council of Canada) 1967 Science and Engineering Scholarship for Graduate Education
- 1983 - 1985 Sir James Lougheed Award of Distinction
- 1982 - 1983 Society of Chemical Industry Merit Award
- 1982 - 1983 Lieutenant Governor's Gold Medal - Highest Academic Achievement in graduating class of the Faculty of Science (fix tabs)
- 1978 - 1979 Highest Honors Standing in Grade XII in the Edmonton Public School System

### **Outreach and Mentoring of High School Students and Undergraduates**

Through the Canada Gairdner Foundation I have travelled extensively throughout Canada giving science lectures to high school students and meeting with them to discuss why science is something that they may wish to consider career-wise. The lectures describe simple concepts of biophysics and how physics can inform on how biomolecules function. Independently I mentor a series of high school students and undergraduates that I have met through these lectures, keeping in touch electronically. I discuss careers in science with them and assist with selection of courses that would best meet their needs and satisfy their interests. A sampling of cities where I have met with high school students includes: Lethbridge, Edmonton, Calgary, Winnipeg, Montreal, St. John's, Toronto, Ottawa, Thunder Bay, Charlottetown, Halifax, Fredricton.

In the past 12 months I have been a speaker at:

- (1) StAR (Student Advancement Research), which is a paid 6-week internship program for Black, Indigenous, and Filipino students to get some lab experience.
- (2) SSuRe (SickKids Summer Undergraduate Research), which is a paid 3-month internship for students to gain research insights.
- (3) Toronto local high schools
- (4) Exploring by the Seat of your Pants, "Unlocking the Mysteries of Life with Dr. Lewis Kay"

## Publications

1. J. Gariépy, Lewis E. Kay, I.D. Kuntz, B.D. Sykes and R.S. Hodges. Nuclear Magnetic Resonance Determination of Metal-Proton Distances in a Synthetic Calcium Binding Site of Rabbit Skeletal Troponin-C. *Biochemistry* **24**, 544-550 (1985). doi:10.1021/bi00323a045.
2. D.C. Corson, T.C. Williams, Lewis E. Kay and B.D. Sykes. <sup>1</sup>H NMR Spectroscopic Studies of Calcium Binding Proteins: 1: Stepwise Proteolysis of the C-Terminal  $\alpha$ -Helix of a Helix-Loop-Helix Metal-Ion-Binding Domain. *Biochemistry* **25**, 1817-1826 (1986). doi:10.1021/bi00355a055.
3. Lewis E. Kay and J.H. Prestegard. An Application of Pulse-Gradient Double Quantum Spin Echoes to Diffusion Measurements on Molecules with Scalar-Coupled Spins. *J. Magn. Reson.* **67**, 103-113 (1986). doi:10.1016/0022-2364(86)90413-0.
4. Lewis E. Kay, J.N. Scarsdale, D.R. Hare and J.H. Prestegard. Simulation of Two-Dimensional Cross-Relaxation Spectra in Strongly Coupled Spin Systems. *J. Magn. Reson.* **68**, 515-525 (1986). doi:10.1016/0022-2364(86)90340-9.
5. Lewis E. Kay, T.A. Holak, B.A. Johnson, I.M. Armitage and J.H. Prestegard. Second-Order Effects in Two-Dimensional Cross-Relaxation Spectra of Proteins: Investigation of Glycine Spin Systems. *J. Am. Chem. Soc.* **108**, 4242-4244 (1986). doi:10.1021/ja00274a076.
6. Lewis E. Kay, M. Pascone, B.D. Sykes and J.W. Shriver. <sup>19</sup>F Nuclear Magnetic Resonance as a Probe of Structural Transitions and Cooperative Interactions in Heavy Meromyosin. *J. Biol. Chem.* **262**, 1984-1988 (1987). doi:10.1016/S0021-9258(18)61608-4.
7. Lewis E. Kay and J.H. Prestegard. Methyl Group Dynamics from Relaxation of Double Quantum Filtered NMR Signals: Application to Deoxycholate. *J. Am. Chem. Soc.* **109**, 3829-3835 (1987). doi:10.1021/ja00247a002.
8. Lewis E. Kay, P.-J. Jones and J.H. Prestegard. Strong Coupling Effects in the Homonuclear RELAY Experiment, with Applications to Leucine Spin Systems of Octanoyl-Acyl Carrier Protein. *J. Magn. Reson.* **72**, 392-396 (1987). doi:10.1016/0022-2364(87)90307-6.
9. Lewis E. Kay, T.L. Jue, B. Bangerter and P.C. Demou. Sensitivity Enhancement of <sup>13</sup>C T<sub>1</sub> Measurements Via Polarization Transfer. *J. Magn. Reson.* **73**, 558-564 (1987). doi:10.1016/0022-2364(87)90024-2.
10. Lewis E. Kay and R.E.D. McClung. A Product Operator Description of AB and ABX Spin Systems. *J. Magn. Reson.* **77**, 258-273 (1988). doi:10.1016/0022-2364(88)90177-1.
11. Lewis E. Kay, T.A. Holak and J.H. Prestegard. AX<sub>3</sub> Spin System Dynamics from Forbidden Cross Peak Intensities in Double Quantum Spectra, with Application to Acyl Carrier Protein. *J. Magn. Reson.* **76**, 30-40 (1988). doi:10.1016/0022-2364(88)90198-9.

12. Lewis E. Kay and J.H. Prestegard. Spin-Lattice Relaxation Rates of Coupled Spins from 2D Accordion Spectroscopy. *J. Magn. Reson.* **77**, 599-605 (1988). doi:10.1016/0022-2364(88)90021-2.
13. Lewis E. Kay and J.H. Prestegard. Simultaneous Measurement of  $^{13}\text{C}$  Multiplicities and  $^1\text{H}$  and  $^{13}\text{C}$  Chemical Shifts. *J. Magn. Reson.* **78**, 172-177 (1988). doi:10.1016/0022-2364(88)90170-9.
14. A.F. Frederick, Lewis E. Kay and J.H. Prestegard. Location of Divalent Ion Sites in Acyl Carrier Protein Using Relaxation Perturbed 2D NMR. *FEBS Lett.* **238**, 43-48 (1988). doi:10.1016/0014-5793(88)80222-9.
15. D.M. LeMaster, Lewis E. Kay, A.T. Brünger and J.H. Prestegard. Protein Dynamics and Distance Determination by NOE Measurements. *FEBS Lett.* **236**, 71-76 (1988). doi:10.1016/0014-5793(88)80287-4.
16. Lewis E. Kay, D.S. Thomson and J.H. Prestegard. Extraction of  $^1\text{H}$ - $^1\text{H}$  and  $^1\text{H}$ - $^{13}\text{C}$  Dipolar Couplings from Spectra Acquired in Inhomogeneous Magnetic Fields. *Magn. Reson. Chem.* **26**, 860-866 (1988). doi:10.1002/mrc.1260261010.
17. A. Bax, Lewis E. Kay, S.W. Sparks and D.A. Torchia. Line Narrowing of Amide Proton Resonances in 2D NMR Spectra of Proteins. *J. Am. Chem. Soc.* **111**, 408-409 (1989). doi:10.1021/ja00183a082.
18. D. Marion, Lewis E. Kay, S.W. Sparks, D.A. Torchia and A. Bax. Three-Dimensional Heteronuclear NMR of  $^{15}\text{N}$  Labeled Proteins. *J. Am. Chem. Soc.* **111**, 1515-1517 (1989). doi:10.1021/ja00186a066.
19. Lewis E. Kay, D. Marion and A. Bax. Practical Aspects of 3D Heteronuclear NMR of Proteins. *J. Magn. Reson.* **84**, 72-84 (1989). doi:10.1016/0022-2364(89)90006-1.
20. Lewis E. Kay and A. Bax. Separation of NH and  $\text{NH}_2$  Resonances in  $^1\text{H}$ -Detected Heteronuclear Multiple-Quantum Correlation Spectra. *J. Magn. Reson.* **84**, 598-603 (1989). doi:10.1016/0022-2364(89)90125-X.
21. Lewis E. Kay, B. Brooks, S.W. Sparks, D.A. Torchia and A. Bax. Measurement of NH-C $\alpha$ H Coupling Constants in Staphylococcal Nuclease by Two-Dimensional NMR and Comparison with X-ray Crystallographic Results. *J. Am. Chem. Soc.* **111**, 5488-5490 (1989). doi:10.1021/ja00196a078.
22. D. Marion, P.C. Driscoll, Lewis E. Kay, P.T. Wingfield, A. Bax, A.M. Gronenborn and G.M. Clore. Overcoming the Overlap Problem in the Assignment of  $^1\text{H}$  NMR Spectra of Larger Proteins by Use of Three-Dimensional Heteronuclear  $^1\text{H}$ - $^{15}\text{N}$  Hartmann-Hahn-Multiple Quantum Coherence and Nuclear Overhauser-Multiple Quantum Coherence Spectroscopy: Application to Interleukin 1 $\beta$ . *Biochemistry* **28**, 6150-6156 (1989). doi:10.1021/bi00441a004.

23. Lewis E. Kay, D.A. Torchia and A. Bax. Backbone Dynamics of Proteins as Studied by  $^{15}\text{N}$  Inverse Detected Heteronuclear NMR Spectroscopy: Application to Staphylococcal Nuclease. *Biochemistry* **28**, 8972-8979 (1989). doi:10.1021/bi00449a003.
24. Lewis E. Kay and A. Bax. New Methods for the Measurement of NH-C $\alpha$ H Coupling Constants of  $^{15}\text{N}$ -Labeled Proteins. *J. Magn. Reson.* **86**, 110-126 (1990). doi:10.1016/0022-2364(90)90215-U.
25. J.D. Forman-Kay, A.M. Gronenborn, Lewis E. Kay, P.T. Wingfield and G.M. Clore. Studies on the Solution Conformation of Human Thioredoxin using Heteronuclear  $^{15}\text{N}$ - $^1\text{H}$  Nuclear Magnetic Resonance Spectroscopy. *Biochemistry* **29**, 1566-1572 (1990). doi:10.1021/bi00458a030.
26. A. Bax, M. Ikura, Lewis E. Kay, D.A. Torchia and R. Tschudin. Comparison of Different Modes of Two-Dimensional Reverse-Correlation NMR for the Study of Proteins. *J. Magn. Reson.* **86**, 304-318 (1990). doi:10.1016/0022-2364(90)90262-8.
27. M. Ikura, Lewis E. Kay, R. Tschudin and A. Bax. Three-Dimensional NOESY-HMQC Spectroscopy of a  $^{13}\text{C}$ -Labeled Protein. *J. Magn. Reson.* **86**, 204-209 (1990). doi:10.1016/0022-2364(90)90227-Z.
28. Lewis E. Kay, M. Ikura and A. Bax. Proton-Proton Correlation via Carbon-Carbon Couplings: A Three-Dimensional NMR Approach for the Assignment of Aliphatic Resonances in Proteins Labeled with Carbon-13. *J. Am. Chem. Soc.* **112**, 888-889 (1990). doi:10.1021/ja00158a070.
29. D.A. Torchia, S.W. Sparks, H.B.R. Cole, D.M. Baldisseri, Lewis E. Kay, D. Marion and A. Bax. Assignments, Structure and Dynamics of Staphylococcal Nuclease. *Polymer Preprints* **91**, 2618-2619 (1990).
30. M. Ikura, Lewis E. Kay, D.A. Torchia, C. Klee and A. Bax. Novel Approaches for Obtaining Resonance Assignments of Larger Proteins. *Polymer Preprints* **91**, (1990).
31. M. Ikura, D. Marion, Lewis E. Kay, H. Shih, M. Krinks, C.B. Klee and A. Bax. Heteronuclear 3D NMR and Isotopic Labeling of Calmodulin: Towards the Complete Assignment of the  $^1\text{H}$  NMR Spectrum. *Biochem. Pharmacol.* **40**, 153-160 (1990). doi:10.1016/0006-2952(90)90190-V.
32. M. Ikura, Lewis E. Kay and A. Bax. A Novel Approach for Sequential Assignment of  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{15}\text{N}$  Spectra of Larger Proteins: Heteronuclear Triple Resonance 3D NMR Spectroscopy. Application to Calmodulin. *Biochemistry* **29**, 4659-4667 (1990). doi:10.1021/bi00471a022.
33. A. Bax, G.M. Clore, P.C. Driscoll, A.M. Gronenborn, P.C. Driscoll, M. Ikura and Lewis E. Kay. Practical Aspects of Proton-Carbon-Carbon-Proton Three-Dimensional Correlation Spectroscopy of  $^{13}\text{C}$  Labeled Proteins. *J. Magn. Reson.* **87**, 620-627 (1990). doi.org:10.1016/0022-2364(90)90320-9.

34. G.M. Clore, A. Szabo, A. Bax, Lewis E. Kay, P.C. Driscoll and A.M. Gronenborn. Deviations from the Simple Two Parameter Model-Free Approach to the Interpretation of Nitrogen-15 Nuclear Magnetic Relaxation of Proteins. *J. Am. Chem. Soc.* **112**, 4989-4991 (1990). doi:10.1021/ja00168a070.
35. Lewis E. Kay, M. Ikura, R. Tschudin and A. Bax. Three-Dimensional Triple-Resonance NMR Spectroscopy of Isotopically Labelled Proteins. *J. Magn. Reson.* **89**, 496-514 (1990). doi:10.1016/j.jmr.2011.09.004.
36. Lewis E. Kay, G.M. Clore, A. Bax and A.M. Gronenborn. Four-Dimensional Heteronuclear Triple-Resonance Spectroscopy of a Protein in Solution. Application to Interleukin-1 $\beta$  in Solution. *Science* **249**, 411-414 (1990). doi:10.1126/science.2377896.
37. G.M. Clore, Lewis E. Kay, A. Bax and A.M. Gronenborn. Four-Dimensional  $^{13}\text{C}/^{13}\text{C}$ -Edited Nuclear Overhauser Enhancement Spectroscopy of a Protein in Solution: Application to Interleukin-1 $\beta$ . *Biochemistry* **30**, 12-18 (1991). doi.org:10.1021/bi00215a002.
38. A. Bax, M. Ikura, Lewis E. Kay and G. Zhu. Removal of  $F_1$  Baseline Distortion and Optimization of Folding in Multidimensional NMR Spectra. *J. Magn. Reson.* **91**, 174-178 (1991). doi:10.1016/0022-2364(91)90422-P.
39. Lewis E. Kay, M. Ikura and A. Bax. The Design and Optimization of Complex NMR Experiments: Application to a Triple-Resonance Pulse Scheme Correlating  $\text{H}\alpha$ , NH and  $^{15}\text{N}$  Chemical Shifts in  $^{15}\text{N}$ - $^{13}\text{C}$  Labeled Proteins. *J. Magn. Reson.* **91**, 84-92 (1991). doi:10.1016/0022-2364(91)90410-U.
40. Lewis E. Kay, M. Ikura, G. Zhu and A. Bax. Four-Dimensional Heteronuclear Triple-Resonance NMR of Isotopically Enriched Proteins for Sequential Assignment of Backbone Atoms. *J. Magn. Reson.* **91**, 422-428 (1991). doi:10.1016/0022-2364(91)90208-B.
41. R. Boelens, C. Griesinger, Lewis E. Kay, D. Marion and E.R.P. Zuiderweg. Applicability and Limitations of Three-Dimensional NMR Spectroscopy for the Study of Proteins in Solution. In "Computational Aspects of the Study of Biological Macromolecules." Vol 225, pp 127-150, NATO ASI Series, Eds. J.C. Hoch, F.M. Poulsen, C. Redfield; Springer, Boston, MA (1991). doi:10.1007/978-1-4757-9794-7\_9.
42. Lewis E. Kay, J.D. Forman-Kay, W.D. McCubbin and C.M. Kay. Solution Structure of a Polypeptide Dimer Comprising the Fourth  $\text{Ca}^{2+}$ -Binding Site of Troponin C by Nuclear Magnetic Resonance Spectroscopy. *Biochemistry* **30**, 4323-4333 (1991). doi:10.1021/bi00231a031.
43. M. Ikura, Lewis E. Kay, M. Krinks and A. Bax. A Triple-Resonance Multidimensional NMR Study of Calmodulin Complexed with the Binding Domain of Myosin Light-Chain Kinase: Indication of a Conformational Change in the Central Helix. *Biochemistry* **30**, 5498-5504 (1991). doi:10.1021/bi00236a024.

44. Lewis E. Kay and D.A. Torchia. The Effects of Dipolar Cross Correlation on  $^{13}\text{C}$  Methyl-Carbon  $T_1$ ,  $T_2$  and NOE Measurements in Macromolecules. *J. Magn. Reson.* **95**, 536-547 (1991). doi:10.1016/0022-2364(91)90167-R.
45. M. Ikura, S. Spera, G. Barbato, Lewis E. Kay, M. Krinks and A. Bax. Secondary Structure and Side-Chain  $^1\text{H}$  and  $^{13}\text{C}$  Resonance Assignments of Calmodulin by Heteronuclear Multidimensional NMR Spectroscopy. *Biochemistry* **30**, 9216-9228 (1991). doi.org:10.1021/bi00102a013.
46. M. Ikura, Lewis E. Kay and A. Bax. Improved Three-Dimensional  $^1\text{H}$ - $^{13}\text{C}$ - $^1\text{H}$  Correlation Spectroscopy of a  $^{13}\text{C}$  Labeled Protein Using Constant-Time Evolution. *J. Biomol. NMR.* **1**, 299-304, (1991). doi.org:10.1007/bf01875522.
47. A. Bax, M. Ikura, Lewis E. Kay, G. Barbato and S. Spera. Multidimensional Triple Resonance NMR Spectroscopy of Isotopically Uniformly Enriched Proteins: A Powerful New Strategy for Structure Determination. In "Protein Conformation." pp 108-134, Ciba Foundation Symposium 161, Eds. D.J. Chadwick, K. Widdows, Wiley, Chichester (1991). doi:10.1002/9780470514146.ch8.
48. L.K. Nicholson, Lewis E. Kay, D.M. Baldisseri, J. Arango, P.E. Young, A. Bax and D.A. Torchia. Dynamics of Methyl Groups in Proteins as Studied by Proton-Detected  $^{13}\text{C}$  NMR Spectroscopy. Application to the Leucine Residues of Staphylococcal Nuclease. *Biochemistry* **31**, 5253-5263 (1992). doi:10.1021/bi00138a003.
49. Lewis E. Kay, L.K. Nicholson, F. Delaglio, A. Bax and D.A. Torchia. Pulse Sequences for Removal of the Effects of Cross Correlation Between Dipolar and Chemical-Shift Anisotropy Relaxation Mechanisms on the Measurement of Heteronuclear  $T_1$  and  $T_2$  Values in Proteins. *J. Magn. Reson.* **97**, 359-375 (1992). doi:10.1016/0022-2364(92)90320-7.
50. G. Barbato, M. Ikura, Lewis E. Kay, R.W. Pastor and A. Bax. Backbone Dynamics of Calmodulin Studied by  $^{15}\text{N}$  Relaxation Using Inverse Detected Two-Dimensional NMR Spectroscopy: The Central Helix is Flexible. *Biochemistry* **31**, 5269-5278 (1992). doi:10.1021/bi00138a005.
51. Lewis E. Kay, T.E. Bull, L.K. Nicholson, C. Griesinger, H. Scwalbe, A. Bax and D.A. Torchia. The Measurement of Heteronuclear Transverse Relaxation Times in  $\text{AX}_3$  Spin Systems via Polarization Transfer Techniques. *J. Magn. Reson.* **100**, 538-558 (1992). doi:10.1016/0022-2364(92)90058-F.
52. Lewis E. Kay and T.E. Bull. Heteronuclear Transverse Relaxation in  $\text{AMX}$ ,  $\text{AX}_2$  and  $\text{AX}_3$  Spin Systems. *J. Magn. Reson.* **99**, 615-622 (1992). doi:10.1016/0022-2364(92)90218-V.
53. Lewis E. Kay, M. Wittekind, M.A. McCoy, M.S. Friedrichs and L. Mueller. 4D Triple-Resonance Experiments for Assignment of Protein Backbone Nuclei Using Constant-Time Evolution Periods. *J. Magn. Reson.* **98**, 443-450 (1992). doi:10.1016/0022-2364(92)90146-X.

54. Lewis E. Kay, M. Ikura, A.A. Grey and D.R. Muhandiram. Three-Dimensional NMR Experiments for the Separation of Side-Chain Correlations in Proteins via the Carbonyl Chemical Shift. *J. Magn. Reson.* **99**, 652-659 (1992). doi:10.1016/0022-2364(92)90223-T.
55. Lewis E. Kay. A Three-Dimensional NMR Experiment for the Separation of Aliphatic Carbon Chemical Shifts via the Carbonyl Chemical Shift in  $^{15}\text{N}$ ,  $^{13}\text{C}$  Labeled Proteins. *J. Magn. Reson. Series B* **101**, 110-113 (1993). doi:10.1006/jmrb.1993.1018.
56. M. Görlach, M. Wittekind, B.T. Farmer II, Lewis E. Kay and L. Mueller. Measurement of  $^3J_{\text{HN}\alpha}$  Vicinal Coupling Constants in Proteins. *J. Magn. Reson. Series B* **101**, 194-197 (1993). doi:10.1006/jmrb.1993.1031.
57. Lewis E. Kay, P. Keifer and T. Saarinen. Pure Absorption Gradient Enhanced Heteronuclear Single Quantum Correlation Spectroscopy with Improved Sensitivity. *J. Am. Chem. Soc.* **114**, 10663-10665 (1992). doi:10.1021/ja00052a088.
58. D.A. Torchia, L.K. Nicholson, H.B.R. Cole and Lewis E. Kay. Heteronuclear NMR Studies of the Molecular Dynamics of Staphylococcal Nuclease. In "Topics in Molecular and Structural Biology Series: NMR of Proteins." pp 190-219, Eds. G.M. Clore and A.M. Gronenborn; The Macmillan Press Ltd (1993). doi.org:10.1007/978-1-349-12749-8\_7.
59. Lewis E. Kay. A Pulsed-Field Gradient-Enhanced Three-Dimensional NMR Experiment for Correlating  $^{13}\text{C}\alpha/^{13}\text{C}\beta$ ,  $^{13}\text{C}'$ ,  $^1\text{H}\alpha$  Chemical Shifts in Uniformly  $^{13}\text{C}$ -Labeled Proteins Dissolved in  $\text{H}_2\text{O}$ . *J. Am. Chem. Soc.* **115**, 2055-2057 (1993). doi:10.1021/ja00058a072.
60. Lewis E. Kay, G.-Y. Xu, A.U. Singer, D.R. Muhandiram and J.D. Forman-Kay. A Gradient-Enhanced HCCH-TOCSY Experiment for Recording Side-Chain  $^1\text{H}$  and  $^{13}\text{C}$  Correlations in  $\text{H}_2\text{O}$  Samples of Proteins. *J. Magn. Reson. Series B* **101**, 333-337 (1993). doi:10.1006/jmrb.1993.1053.
61. D.R. Muhandiram, G.Y. Xu and Lewis E. Kay. An Enhanced-Sensitivity Pure Absorption Gradient 4D  $^{15}\text{N}$ - $^{13}\text{C}$ -Edited NOESY Experiment. *J. Biol. NMR* **3**, 463-470 (1993). doi:10.1007/BF00176011.
62. D.R. Muhandiram, N.A. Farrow, G.-Y. Xu, S. H. Smallcombe and Lewis E. Kay. A Gradient  $^{13}\text{C}$  NOESY-HSQC Experiment for Recording NOESY Spectra of  $^{13}\text{C}$ -Labeled Proteins Dissolved in  $\text{H}_2\text{O}$ . *J. Magn. Reson. Series B* **102**, 317-321 (1993). doi:10.1006/jmrb.1993.1102.
63. T. Yamazaki, J.D. Forman-Kay and Lewis E. Kay. Two-Dimensional NMR Experiments for Correlating  $^{13}\text{C}\beta$ - $^1\text{H}\delta/\epsilon$  Chemical Shifts of Aromatic Residues in  $^{13}\text{C}$  Labeled Proteins via Scalar Couplings. *J. Am. Chem. Soc.* **115**, 11054-11055 (1993). doi:10.1021/ja00076a099.
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469. T.H. Kim, B.J. Payliss, M.L. Nosella, I.T.W. Lee, Y. Toyama, J.D. Forman-Kay and Lewis E. Kay. Interaction Hot Spots for Phase Separation Revealed by NMR Studies of a CAPRIN1 Condensed Phase. *Proc. Natl. Acad. Sci. USA* **118**, e2104897118. Epub 31 May 2021. doi:10.1073/pnas.2104897118.
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499. J. P. Bonin, J. M. Aramini, Y. Dong, H. Wu, and Lewis E. Kay. AlphaFold2 as a Replacement for Solution NMR Structure Determination of Small Proteins: Not So Fast! 2024 Jul;364:107725. doi: 10.1016/j.jmr.2024.107725. Epub 2024 Jun 19.
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501. A. K. Rangadurai, L. Ruetz, R. Ahmed, K. Lo, M. Tollinger, J. D. Forman-Kay, C. Kreutz and Lewis E. Kay. Phase Separation Modulates the Thermodynamics and Kinetics of RNA Hybridization. *J Am Chem Soc.* 2024 Jul 24;146(29):19686-19689. doi: 10.1021/jacs.4c06530. Epub 2024 Jul 11.
502. B. Yu, N. Bolik-Coulon, A. K. Rangadurai, Lewis E. Kay, J. Iwahara. Gadolinium-Based Spin Relaxation Measurements of Near-Surface Electrostatic Potentials of Biomolecules. *J Am Chem Soc.* 2024 Jul 19;146(30):20788-20801. doi: 10.1021/jacs.4c04433. Epub 2024 Jul 19.
503. R. Ahmed, M. Liang, R. P. Hudson, A. T. Rangadurai, S. K. Huang, J. D. Forman-Kay and Lewis E. Kay. Atomic Resolution Map of the Solvent Interactions Driving SOD1 Unfolding in CAPRIN1 Condensates. *Proc. Natl. Acad. Sci. USA.* 2024 Aug 27;121(35):e2408554121. doi: 10.1073/pnas.2408554121. Epub 22 Aug 2024.

## **TEACHING**

Undergraduate coordinator of Molecular Biophysics BSc program offered through the Department of Physics.

- 1) Chemistry 1455H, NMR Spectroscopy 1: Introduction to NMR: Theory and Application.
- 2) Chemistry 1456H, NMR Spectroscopy 2: Advanced Theory and Application.
- 3) MMG 1012Y Protein Structure and Function.
- 4) MMG 1014Y. Macromolecular Biology. Special Topics Course.
- 5) Biochemistry 2021.
- 6) BCH 2024H Biomolecular Dynamics and Function.

## **ORGANIZATION OF NATIONAL AND INTERNATIONAL MEETINGS**

"Frontiers in Structural Biology Symposium" June 1993.

- 2) "NMR of Biomolecules" for the Canadian Society of Chemistry May 1995.
- 3) Minisymposium on Biomolecular NMR, Canadian Society for Biochemistry and Molecular and Cellular Biology. June 1996.
- 4) International Society of Magnetic Resonance in Biological Systems Conference, Toronto, August 2002.

## **RESEARCH TRAINING**

- 1) Ouwan Zhang, Graduate Student (PhD) in Chemistry with Dr. Julie Forman-Kay, June 1992 - April 1997.
- 2) Neil Farrow, Post-Doctoral Associate, September 1992 - July 1997.
- 3) Steven Pascal, Post-Doctoral Associate, March 1993 - July 1996.
- 4) Toshio Yamazaki, Post-Doctoral Associate, April 1993 - April 1995.
- 5) Yves Aubin, Post-Doctoral Associate, September 1993 - October 1995.
- 6) Mike Rosen, Post-Doctoral Associate, October 1993 - January 1996.

- 7) Ping Xu, Post-Doctoral Associate, January 1994 - September 1994.
- 8) Li-Lin Tay, Graduate Student (MSc) in Chemistry with Dr. Bill Reynolds. September 1994 - November 1996.
- 9) N. Rao, Post-Doctoral Associate, April 1995 - October 1996.
- 10) D. Yang, Post-Doctoral Associate, May 1995 – May 2001.
- 11) Kevin Gardner, Post-Doctoral Associate, August 1995 - July 1998.
- 12) Pascale Legault, Post-Doctoral Associate, September 1995 - July 1998.
- 13) Sebastian Vincent, Post-Doctoral Associate, May 1996 - September 1997.
- 14) Catherine Zwahlen, Post-Doctoral Associate, May 1996 - December 1998.
- 15) Joel Tolman, Post-Doctoral Associate, December 1997 - September 1999.
- 16) Logan Donaldson, Post-Doctoral Associate, July 1998 - September 2000.
- 17) Geoff Mueller, Post-Doctoral Associate, September 1998 – May 2001.
- 18) James Choy, Post-Doctoral Associate, October 1998 – Nov 2005.
- 19) Nikolai Skrynnikov, Post-Doctoral Associate, January 1998 – Sept. 2002.
- 20) Frans Mulder, Post-Doctoral Associate, February 1998 – July 2001.
- 21) Johan Evenäs, Post-Doctoral Associate, July 1999 – February 2001.
- 22) Tony Mittermaier, Graduate Student, September 1997 – Jan 2004.
- 23) Natalie Goto, Graduate Student, September 1997 - February 2001.
- 24) Peter Hwang, Graduate Student, September 1999 – Nov 2005.
- 25) Jason Ollerenshaw, Graduate Student, September 2000 – Sept 2005.
- 26) Vitali Tugarinov, Post-Doctoral Associate, December 2000 – July 2007.
- 27) Oscar Millet, Post-Doctoral Associate, September 2000 – April 2004.
- 28) Dimitry Korzhnev, Post-Doctoral Associate, September 2001-Sept 2010.
- 29) Irina Bezsonova, Graduate Student, September 2001-Sept. 2008

- 30) Karen Kloibler, Post-Doctoral Associate, September 2001-Jan 2004.
- 31) Pramodh Vallurupalli, Post-Doctoral Associate, Oct 2002-June 2009.
- 32) Remco Sprangers, Post-Doctoral Associate, April 2003-March 2008.
- 33) Algirdas Velyvis, Post-Doctoral Associate, April 2003-2009.
- 34) Philipp Neudecker, Post-Doctoral Associate, April 2004-Oct. 2010.
- 35) Jason Malthrop, Graduate Student, Sept. 2004-February 2008.
- 36) Flemming Hansen, Post-Doctoral Associate, Sept 2005-Sept 2010.
- 37) Patrik Lundström, Post-Doctoral Associate, Oct 2005-February 2009.
- 38) Eugene V. Tischenko, Post-Doctoral Associate, Oct 2005-August 2009.
- 39) Tomasz Religa, Post-Doctoral Associate, August 2008-October 2011.
- 40) Andy Baldwin, Post-Doctoral Associate, May 2008-July 2012.
- 41) Amy Ruschak, Post-Doctoral Associate, June 2008-April 2012.
- 42) Michael Latham, Post-Doctoral Associate, July 2008-Nov 2014.
- 43) Hugo van Ingen, Post-Doctoral Associate, August 2008-Dec 2010.
- 44) Guillaume Bouvignies, Post-Doctoral Associate, August 2008-November 2013.
- 45) Julia Barette, Graduate Student, January 2009, August 2011.
- 46) Alex Hansen, Post-doctoral Fellow, January 2009-February 2014.
- 47) Rina Rosenzweig, Post-Doctoral Associate, September 2010-June 2016.
- 48) Lichi Shi, Post-Doctoral Associate, Jan 2011-Sept 2014.
- 49) Julianne Kitevski, Post-Doctoral Associate, July 2013-June 2017.
- 50) Ashok Sekhar, Post-Doctoral Associate, July 2011-August 2017.
- 51) Rafal Augustyniak, Post-Doctoral Associate, May 2012-June 2018.
- 52) Dong Long, Post-Doctoral Associate, Oct 2012-May 2015.

- 53) Anne Schuetz, Post-Doctoral Associate, August 2013-May 2017.
- 54) Enrico Rennella, Post-Doctoral Associate / Senior Research Associate, June 2014-Present.
- 55) Rui Hwang, Post-Doctoral Associate, Sept 2014-May 2020.
- 56) Rob Culik, Post-Doctoral Associate, Sept 2014-August 2017.
- 57) Siavash Vahidi, Post-Doctoral Associate, Sept 2015-May 2020.
- 58) Zev Ripstein, Graduate Student, Sept 2015-Oct 2020
- 59) Tairan Yuwen, Post-Doctoral Associate, Nov 2015-Nov 2019
- 60) Jacob Brady, Post-Doctoral Associate, Nov 2015-Nov 2019.
- 61) Tae Hun Kim, Post-Doctoral Associate, Dec 2016-Dec 2022.
- 62) Gili Abramov, Post-Doctoral Associate, Jan 2018-Feb 2022.
- 63) Rob Harkness, Post-Doctoral Associate, July 2018-March 2024.
- 64) Alex Conicella, Post-Doctoral Associate, July 2018-Feb 2021.
- 65) Leo Wong, Post-Doctoral Associate, November 2018-July 2020.
- 66) T. Reid Alderson, Post-Doctoral Associate, February 2019-Feb 2022.
- 67) Yuki Toyama, Post-Doctoral Associate, April 2019-Sept 2022.
- 68) Zev Ripstein, Post-Doctoral Associate, Oct 2020-Aug 2021.
- 69) Alex Server, Graduate Student, Sept 2020-Present.
- 70) Rashik Ahmed, Post-Doctoral Associate, Nov. 1, 2021-Present.
- 71) Atul Kaushik Rangadurai, Post-Doctoral Associate, Nov. 1, 2021-Aug 2024
- 72) Nicolas BolikCoulon, Post-Doctoral Associate, Dec. 1, 2021-Present.
- 73) Jeff Paul Bonin, Post-Doctoral Associate, July 1, 2022-Present.
- 74) Philipp Roessler, Post-Doctoral Associate, Jan 1, 2023 – Present.
- 75) Aidan Estelle, Post-Doctoral Associate, April 15, 2023-Present.

Current statistics show that only about 10% of trainees go on to independent research careers – yet 40 of 57 trainees who have left the Kay laboratory are on faculty at major research-intensive universities, an additional 5 are scientists in government laboratories and 6 are in industry. My trainees are now professors at University of Toronto, Université de Montréal, McGill University, University of Ottawa, York University, Guelph University, University of Manitoba, and other academic centers, group leaders at Health Canada and industrial sites. Other trainees have established careers at universities throughout the U.S., Europe, Israel and Asia at sites as diverse as Singapore, Dallas, New York, London and Hyderabad. Included in this list are a very significant number of women.

Listed below are former trainees that are now on faculty across the world: Steven Pascal, Professor, Massey University, New Zealand; Toshio Yamazaki, Professor, Osaka University, Japan; Mike Rosen, Professor, Southwestern Medical Center and Howard Hughes Investigator, USA; Daiwen Yang, Professor, National University of Singapore, Singapore; Kevin Gardner, Professor, Southwestern Medical Center, USA; Pascale Legault, Professor, University of Montreal, Canada; Joel Tolman, Professor, Johns Hopkins University, USA; Logan Donaldson, Professor, York University, Canada; James Choy, Professor, University of Western Ontario, Canada; Nikolai Skrynnikov, Professor, Purdue University, USA; Frans Mulder, Professor, University of Groningen, Netherlands; Tony Mittermaier, Professor, McGill University, Canada; Natalie Goto, Professor, University of Ottawa, Canada; Oscar Millet, Professor, University of Bilbao, Spain; Voula Kanelis, Professor, University of Toronto, Canada; Remco Sprangers, Group Leader, Max Planck, Tuebingen, Germany; Patrik Lundstrum, Professor, Linköping University, Sweden; Vitali Turgarinov, Professor, University of Maryland, USA; Flemming Hansen, Professor, Imperial College, London, UK; Dmitry Korzhnev, Professor, University of Connecticut; Tomasz Religa, Professor, Case Western Reserve, Cleveland, USA; Amy Ruschak, Professor, Case Western Reserve, Cleveland, USA; Philipp Neudecker, Professor, Jülich Institutes; Pramodh Vallurupalli, Professor, Hyderabad, India; Andy Baldwin, Professor Oxford, Alex Hansen, Research Professor Ohio State, USA; Guillaume Bouvignies, Assistant Professor, CNRS Grenoble France; Irina Bezsonova, Professor, University of Connecticut; Don Long, Professor, University of Science and Technology, China; Michael Latham, Professor, Texas Tech University; Peter Hwang, Professor, University of Alberta; Rina Rosenzweig, Professor Weizmann Institute, Israel; Ashok Sekhar, Professor, Indian Institute of Science, Bengaluru, India; Rafael Augustyniak, Assistant Professor, University of Warsaw, Poland; Anne Schuetz, Professor, University of Munich, Germany; Tairan Yuwen, Assistant Professor, Peking University, China; Rui Hwang, Assistant Professor, University of Guelph, Canada; Siavash Vahidi, Assistant Professor, University of Guelph; Leo Wong, Shenzhen Institute of Advanced Technology (SIAT) in Shenzhen, China; Zev Ripstein, Assistant Professor, University of Manitoba, Canada. Reid Alderson, Assistant Professor, University of Graz, Austria. Tae Hun Kim, Assistant Professor, Case Western Reserve University, Canada. Robert Harkness, Assistant Professor, University of Guelph, Canada. Several trainees have pursued industry related careers: Yves Aubin, Merck Frosst, Canada; Neil Farrow, Beringer-Manheim, USA; Sebastian Vincent, Nestle, Switzerland; Johan Evenas, Astra Zeneca, Sweden as well as independent positions in government laboratories: Ping Xu, NRC, Canada; Geoff Mueller, National Cancer Institute, North Carolina, USA. Eugene Tishchenko, JEOL, USA.

## RESEARCH INTERESTS

Research focuses on the development of NMR techniques for studying macromolecular structure and dynamics and the application of NMR techniques to problems of biological and clinical importance. In particular the research is divided into the following areas:

1) *Methodological Developments*. Research is focused on developing  $^{15}\text{N}$ ,  $^{13}\text{C}$ ,  $^2\text{H}$  multi-dimensional NMR spectroscopy and gradient enhanced spectroscopy to increase the molecular weight limitations currently imposed on protein structure determination using conventional techniques. A second area relates to the development of  $^{15}\text{N}$  and  $^{13}\text{C}$  relaxation techniques for the study of protein dynamics in solution.

2) *Dynamics Studies of Supra-molecular Complexes*. In particular, we are studying the proteasome as well as protein machines involved in disaggregation and the nucleosome. The work builds on our methodological developments that facilitate studies of protein complexes in the MDa molecular weight range.

3) *Protein Dynamics*. Methods are developed and applied to study backbone and sidechain protein dynamics and how dynamic properties change upon ligand binding or folding. Relationships between dynamics and thermodynamics are being developed and applied to binding and folding events.

4) *Protein Folding*. Methods are developed and applied to study protein folding and misfolding, residual structure in unfolded and partially folded states and dynamic properties of these molecules.

5) *Studies of Excited State Protein Conformers*. We are developing the framework to study sparsely populated, transiently formed conformations of proteins that are implicated in function and in disease. The methodology is applied to a wide spectrum of protein systems.